

**UNIVERSIDADE FEDERAL DE PERNAMBUCO
CENTRO DE CIÊNCIAS BIOLÓGICAS
MESTRADO EM BIOQUÍMICA E FISIOLOGIA**

JOSÉ OLIVÁ APOLINÁRIO SEGUNDO

**EFEITO DO TRATAMENTO COM NOVOS DERIVADOS IMIDAZOLIDÍNICOS E
PRAZIQUANTEL SOBRE OS LIPÍDIOS DE CAMUNDONGOS INFECTADOS POR
*SCHISTOSOMA MANSONI***

RECIFE, 2008

JOSÉ OLIVÁ APOLINÁRIO SEGUNDO

**EFEITO DO TRATAMENTO COM NOVOS DERIVADOS IMIDAZOLIDÍNICOS E
PRAZIQUANTEL SOBRE OS LIPÍDIOS DE CAMUNDONGOS INFECTADOS POR
*SCHISTOSOMA MANSONI***

Dissertação apresentada ao Programa de
Pós-graduação em Bioquímica e Fisiologia
como requisito final para obtenção do grau
de Mestre em Bioquímica e Fisiologia pela
Universidade Federal de Pernambuco.

Orientadora: Profa. Dra. Vera Lúcia de
Menezes Lima

Co-orientadora: Profa. Dra. Suely Lins
Galdino

RECIFE, 2008

Apolinário Segundo, José Olivá

Efeito do tratamento com novos derivados imidazolidínicos e praziquantel sobre os lipídios de camudongos infectados por *Schistosoma mansoni* / José Olivá Apolinário Segundo. – Recife: O Autor, 2008.

64 folhas : il., fig., tab.

Dissertação (mestrado) – Universidade Federal de Pernambuco. CCB. Bioquímica e Fisiologia, 2008.

Inclui bibliografia e anexos.

1. Parasitologia 2. Esquistossomose mansônica 3. Colesterol total 4. Farmacologia – imidazolidinas I. Título.

576.89
616.96

CDU (2.ed.)
CDD (22.ed.)

UFPE
CCB – 2008-008

JOSÉ OLIVÁ APOLINÁRIO SEGUNDO

**EFEITO DO TRATAMENTO COM NOVOS DERIVADOS IMIDAZOLIDÍNICOS E
PRAZIQUANTEL SOBRE OS LIPÍDIOS DE CAMUNDONGOS INFECTADOS POR
*SCHISTOSOMA MANSONI***

Dissertação apresentada ao Programa de
Pós-graduação em Bioquímica e Fisiologia
como requisito final para obtenção do grau
de Mestre em Bioquímica e Fisiologia pela
Universidade Federal de Pernambuco.

Defendida em 25 de Fevereiro de 2008.

BANCA EXAMINADORA:

Professora Dra. Patrícia Maria Guedes Paiva
Departamento de Bioquímica - UFPE (Membro interno)

Professora Dra. Luana Cassandra Breitenbach Barroso Coelho
Departamento de Bioquímica - UFPE (Membro interno)

Professora Dra. Mônica Camelo Pessoa de Azevedo Albuquerque
Departamento de Medicina Tropical - UFPE (Membro externo)

Professora Dra. Vera Lúcia de Menezes Lima
Departamento de Bioquímica – UFPE (Presidente)

Ata da defesa de dissertação do Mestrando **José Olivá Apolinário Segundo**, realizada em 25 de fevereiro de 2008, como requisito final para obtenção do título de Mestre em Bioquímica e Fisiologia da UFPE.

Às 09:40 horas, do dia vinte e cinco de fevereiro de 2008, foi aberto no Auditório Prof. Marcionilo Lins – Depto. de Bioquímica, do Centro de Ciências Biológicas da Universidade Federal de Pernambuco o ato de defesa de dissertação do mestrando **José Olivá Apolinário Segundo**, aluno do Curso de Mestrado em Bioquímica e Fisiologia/CCB/UFPE. Iniciando os trabalhos a Profa. Dra. **Vera Lúcia de Menezes** fez a apresentação do aluno, sua orientadora, ela própria, sua co-orientadora Profa. Dra. Suely Lins Galdino, bem como da Banca Examinadora composta por ela própria, na qualidade de Presidente e as professoras doutoras: Patrícia Maria Guedes Paiva, Luana Cassandra Breitenbach Barroso Coelho, ambas do Depto. de Bioquímica/UFPE, e Mônica Camelo Pessôa de Azevedo Albuquerque, do Depto de Medicina Tropical/UFPE. Após as apresentações, a Profa. Dra. Vera Lúcia de Menezes Lima convidou o aluno para a apresentação de sua dissertação intitulada: “**Efeito do Tratamento com Derivados Imidazolidínicos 3-benzil-5-(4- cloro-arylazo)-4-tioxo-imidazolidin-2-oná (LPSF-PT5), 3-(4-cloro-benzil)-5(4-nitro-benzilideno)-imidazolidina-2, 4-diona (LPSF-FZ4) e Praziquantel sobre os Lipídeos de Camundongos Infectados por Schistosoma mansoni**”, e informou que de acordo com o Regimento Interno do Curso, o candidato dispõe de até 50 (cinquenta) minutos para apresentação do trabalho e o tempo de argüição para cada examinador, juntamente com o tempo gasto pelo aluno para responder às perguntas será de 30 (trinta) minutos. O aluno procedeu à explanação e comentários acerca do tema em **35 (trinta e cinco) minutos**. Após a apresentação do mestrando, a Sra. Presidente convidou a Banca Examinadora para ocupar seus lugares e passou a palavra a primeira examinadora, Profa. Dra. Mônica Camelo Pessôa de Azevedo Albuquerque que agradeceu o convite, fez alguns comentários e sugestões, e iniciou sua argüição. Ao final, a referida professora deu-se por satisfeita. Em seguida, a Sra. Presidente passou a palavra para a Profa. Dra. Patrícia Maria Guedes Paiva, que agradeceu o convite, fez alguns comentários e sugestões, e iniciou sua argüição. Ao final, a referida professora deu-se por satisfeita. Logo após, a Sra. Presidente passou a palavra para a Profa. Dra. Luana Cassandra Breitenbach Barroso Coelho, que agradeceu ao convite, fez alguns comentários e sugestões, iniciando sua argüição. Ao final, a referida professora deu-se por satisfeita. Em seguida, a Sra. Presidente usou da palavra, na qualidade de orientadora, para tecer alguns comentários, agradecer à Banca Examinadora e parabenizar o candidato. Dando prosseguimento, a sessão foi suspensa para proceder ao julgamento pela Banca Examinadora, a qual se reuniu na Secretaria do Curso. Após alguns comentários, a Banca decidiu, por unanimidade, conceder a menção “**Aprovado com Distinção**”. Nada mais havendo a tratar, lavrei a presente ata que vai assinada por mim, Secretário, e demais membros da Banca Examinadora. Recife, 25 de fevereiro de 2008.

José Olivá Segundo
Mônica C. P. Albuquerque
Patrícia Paiva Guedes Paiva
Vera Lúcia d. L.ia.

AGRADECIMENTOS

Agradeço primeiramente a Deus, por me ajudar nos momentos mais difíceis, por me dar coragem para correr atrás das coisas que acreditei e perseverança para com aquilo que julgava não ter solução.

Agradeço aos meus familiares por me apoiarem, em particular meus pais, por me apoiarem e incentivarem não somente durante o mestrado, mas também durante toda minha vida.

A minha querida Marcela, por me ouvir, apoiar, estimular e cobrar por quase um ano (a distancia). Agradeço também sua compreensão nos momentos que foram necessários, os quais nos serão retribuídos em triplo.

A minha estimada professora-orientadora Vera Lúcia de Menezes Lima. Obrigado pela confiança que me foi depositada. Também obrigado por me orientar e apoiar nas decisões a serem tomadas e executadas durante esta jornada científica.

À Profa. Dra. Suely Lins Galdino por colaborar com os requisitos necessários para a execução deste projeto.

Às Profas. Dras. Maria do Carmo (Nena, como é carinhosamente conhecida) e Mônica Albuquerque por suas colaborações valiosas para que esse trabalho fosse concretizado.

A todos os funcionários do Departamento de Bioquímica da UFPE, em especial Albérico Real, João Virgínio, Djalma Gomes e Sr. Ademar pela disposição em ajudar com boa vontade.

Agradeço a Juliana Kelle, Andréa Apolinário e Fernanda César pelos esforços dedicados para com esta colaboração.

Aos componentes do laboratório de química e metabolismo de lipídios e lipoproteínas pelas ajudas e apoios dispensados. Em particular à Adenor Almeida, Tiago Araújo, Bianka Santana pelos constantes apoios nas horas em que mais precisei.

Também em especial a minha colega de curso Cleideana Bezerra pelos incansáveis apoios e incentivos, desde o início ao término. Também em especial a Aline Farias e Amanda Farias pelos valiosos suportes prestados e amizades adquiridas na reta final do projeto.

Ao CNPQ, CAPES e FINEP por todo o suporte financeiro dado durante a realização deste trabalho.

RESUMO

A Esquistossomose mansônica é a segunda parasitose humana mais prevalente no mundo, sendo no Brasil um problema de saúde pública. A esquistossomose altera o metabolismo lipídico, promovendo diminuição nos níveis de colesterol total (CT), fosfolipídios totais (FT) e triglicerídeos (TG) plasmáticos em hospedeiros como o camundongo. O praziquantel (PZQ) é o principal agente esquistosomicida empregado para combate à doença. Entretanto, o aparecimento de cepas de *Schistosoma mansoni* resistentes ao tratamento convencional, demonstra a necessidade de desenvolvimento de novas drogas esquistosomicidas. Nesse contexto, as imidazolidinas vêm se destacando como fármacos com notória atividade antiparasitária frente ao *S. mansoni* *in vitro* e *in vivo*. Este trabalho teve por objetivo avaliar o efeito do tratamento, nas doses de 50mg/kg/dia e 100mg/kg/dia durante 5 dias, com derivados imidazolidínicos 3-benzil-5-(4-cloro-arilazo)-4-tioxo-imidazolidin-2-ona (LPSF-PT5), 3-(4-cloro-benzil)-5-(4nitro-benzilideno)-imidazolidina-2,4-diona (LPSF-FZ4) e PZQ sobre os lipídios plasmáticos de camundongos infectados e não infectados por *S. Mansoni*. Os resultados mostraram que em camundongos infectados por *S. mansoni*, o tratamento com 100mg/kg/dia de PZQ e LPSF-PT5 reduziu significativamente os níveis plasmáticos de CT em 24.59% e 18.60% respectivamente, bem como os níveis de TG em 31.60% e 31.50%, respectivamente, fenômeno explicável pelo efeito da esquistossomose no animal. Com 50 mg/kg/dia e 100 mg/kg/dia, LPSF-FZ4 promoveu respectivamente, 44.80% e 40.30% de redução nos TG plasmáticos de animais infectados. Já nos animais não infectados, somente foi observada com LPSF-FZ4 diminuição nos TG em 26.7% quando tratados com 50 mg/kg/dia e em 21.7% quando tratados com 100 mg/kg/dia. Portanto, analisando as alterações *in vivo* nos lipídios plasmáticos, o derivado imidazolidínico LPSF-FZ4 representou o fármaco com melhor potencial hipotrigliceridêmico.

Descritores: esquistossomose mansônica, colesterol total, atividade hipotrigliceridemia, praziquantel, derivados imidazolidínicos.

ABSTRACT

The Schistosomiasis mansoni is the second human parasitosis more prevalent in the world, being in Brazil a problem of public health. The schistosomiasis alters lipid metabolism, promoting decrease in plasma levels of total cholesterol (TC), total phospholipids (TF) and triglycerides (TG). The praziquantel (PZQ) is the principal agent schistosomicidal employed for the treatment. However, the apparition of resistant strains to the conventional treatment encourage the development of new anti-schistosomal drugs. In this context, the imidazolidines comes detaching like drugs with potential anti-schistosomal activity front of *S. mansoni* *in vitro* and *in vivo*. This work aimed to evaluated the effect of treatment, at concentrations of 50mg/kg/day and 100mg/kg/day of imidazolidine derivatives 3-benzyl-5-(4-chloro-arylazo)-4-thioxo-imidazolidin-2-one (LPSF-PT5), 3-(4-chloro-benzyl)-5-(4nitro-benzylidene)-imidazolidine-2,4-dione (LPSF-FZ4) and PZQ on lipid metabolism of schistosomiasis mansoni infected and uninfected mice. The results showed that in infected mice, treatment with 100mg/kg/day PZQ and LPSF-PT5 significantly reduced plasma TC levels by 24.59% and by 18.60%, respectively as well TG level by 31.60% e 31.50%, respectively fact possibly justified for schistosomiasis effect in the animal. At dose of 50 mg/kg/day and 100 mg/kg/day, LPSF-FZ4 induced significant decrease by 44.80% and by 40.30% on TG levels from *S. mansoni* mice, respectively. On the other hand, uninfected animals treated with 50 mg/kg/day and 100 mg/kg/day LPSF-FZ4 presented about 26.9% and 21.7% significant decreases on plasma TG level. Therefore, analyzing the alterations *in vivo* of plasma lipids evaluated, imidazolidine derivative LPSF-FZ4 represented the compound with better potential hypotriglyceridemic.

Keywords: schistosomiasis mansoni, total cholesterol, hipotrygliceridemic activity, praziquantel, imidazolidines derivatives

LISTA DE FIGURAS

1. INTRODUÇÃO

Figura 1. Estado de controle da esquistosomose no mundo (ENGELS <i>et al.</i> , 2002).....	11
Figura 2. Distribuição global do <i>S. mansoni</i> (GRYSEELS <i>et al.</i> , 2006)	12
Figura 3 – Ciclo de vida do <i>Schistosoma mansoni</i> (GRYSEELS, et al., 2006).....	14
Figura 4 – Plexo mesentérico de camundongos infectados por <i>S. mansoni</i> , local de acasalamento e oviposição.	15
Figura 5 – Fígado contendo fibroses (pontos brancos) decorrentes de lesões granulomatosas oriundas da infecção por <i>S. mansoni</i>	16
Figura 6 – Estrutura química do praziquantel.....	17
Figura 7 – Estrutura química do anel imidazolidínico.	19
Figura 8 - Estrutura química da imidazolidina-2,4-diona	20
Figura 9 – Estrutura química da 4-Tioxo_imidazolidin-2-oná	20
Figura 10 – Estrutura química do 3-(4-cloro-benzil)-5-(4nitro-benzilideno)-imidazolidina-2,4-diona (LPSF-FZ4).....	21
Figura 11 – Estrutura química do 3-benzil-5-(4-cloro-arylazo)-4-tioxo-imidazolidin-2-oná (LPSF-PT5).....	21
Figura 12 – Estrutura do anel imidazólico.....	23

4. CAPÍTULO I

Figure 1 - Chemical structure of compounds, 3-(4-chloro-benzyl)-5-(4nitro-benzylidene)-imidazolidine-2,4-dione (LPSF-FZ4) (A) and 3-benzyl-5-(4-chloro-arylazo)-4-thioxo-imidazolidin-2-one (LPSF-PT5) (B).	38
--	----

LISTA DE TABELAS

1. INTRODUÇÃO

Tabela 1. Grupo de espécies de Schistosoma (COON, 2005).....	11
Tabela 2 – Algumas características químicas dos compostos 3-(4-cloro-benzil)-5-(4nitro-benzilideno)-imidazolidina-2,4-diona (LPSF-FZ4) e 3-benzil-5-(4-cloro-arylazo)-4-tioxo-imidazolidin-2-ona (LPSF-PT5).....	22

4. CAPÍTULO I

Table 1 - Comparison of biochemical parameters from uninfected and infected mice groups before treatment.....	49
Table 2 - Plasma Total Cholesterol levels before and after 15 days of treatment with praziquantel (PZQ), 3-benzyl-5-(4-chloro-arylazo)-4-thioxo-imidazolidin-2-one (LPSF-PT5) and 3-(4-chloro-benzyl)-5-(4nitro-benzylidene)-imidazolidine-2,4-dione (LPSF-FZ4) in schistosomiasis mansoni infected and uninfected mice.50	
Table 3 - Plasma triglycerides levels before and after 15 days of treatment with praziquantel (PZQ), 3-benzyl-5-(4-chloro-arylazo)-4-thioxo-imidazolidin-2-one (LPSF-PT5) and 3-(4-chloro-benzyl)-5-(4nitro-benzylidene)-imidazolidine-2,4-dione (LPSF-FZ4) in schistosomiasis mansoni infected and uninfected mice.51	
Table 4 - Plasma aspartate aminotransferase activity before and after 15 days of treatment with praziquantel (PZQ), 3-benzyl-5-(4-chloro-arylazo)-4-thioxo-imidazolidin-2-one (LPSF-PT5) and 3-(4-chloro-benzyl)-5-(4nitro-benzylidene)-imidazolidine-2,4-dione (LPSF-FZ4) in schistosomiasis mansoni infected and uninfected mice.....52	
Table 5 - Plasma alanine aminotransferase activities before and after 15 days of treatment with praziquantel (PZQ), 3-benzyl-5-(4-chloro-arylazo)-4-thioxo-imidazolidin-2-one (LPSF-PT5) and 3-(4-chloro-benzyl)-5-(4nitro-benzylidene)-imidazolidine-2,4-dione (LPSF-FZ4) in schistosomiasis mansoni infected and uninfected mice.....53	

SUMÁRIO

1.	INTRODUÇÃO	10
2.	OBJETIVOS.....	26
2.1.	Geral.....	26
2.2.	Específicos	26
3.	REFERÊNCIAS BIBLIOGRÁFICAS.....	27
4.	CAPÍTULO I - EFFECT OF TREATMENT WITH NEW IMIDAZOLIDINE DERIVATIVES AND PRAZIQUANTEL ON PLASMA LIPID OF MICE INFECTED BY <i>SCHISTOSOMA MANSONI</i>.....	34
4.1.	Abstract	36
4.2.	Introduction.....	37
4.3.	Materials and Methods	38
4.4.	Results	40
4.5	Discussion.....	41
4.6.	Acknowledgements	43
4.7.	References.....	43
4.8.	Tables	49
4.9.	Figure legends	54
5.	CONCLUSÕES	55
6.	ANEXOS.....	56

1. INTRODUÇÃO

No ano de 1851, na cidade do Cairo, Egito, Theodor Maximilian Bilharz, anatomicista e helmintologista alemão, descreveu o aparecimento do primeiro esquistossômulo em humanos, caracterizado como um trematoda, inicialmente descrito como *Distomum haematoeum* e, hoje *Schistosoma haematoeum*. Ao mesmo tempo, no Japão, outros cientistas também descreveram o aparecimento de outro trematoda, o *Schistosoma japonicum*. Até 1906 apenas tinha-se conhecimento dessas duas espécies, com suas morfologias descritas. Contudo, em 1907, na Inglaterra, observou-se, ocasionalmente, na urina de pacientes com esquistossomose, o aparecimento de ovos com espículos laterais, os quais não se tinham descrição. Luigi Sambon, da Escola de Medicina Tropical de Londres, tinha descoberto outro trematoda denominado *Schistosoma mansoni* (COON, 2005).

A esquistossomose é uma parasitose causada por helmintos do gênero *Schistosoma*. Estima-se que 200 milhões de pessoas em todo o mundo estejam infectadas pelas diferentes espécies do parasita (BLANCHARD, 2004). Destes, 120 milhões são pacientes sintomáticos da fase aguda e 20 milhões são portadores da fase mais severa da doença (CHITSULO *et al.*, 2000). De acordo com ENGELS *et al.* (2002), ela é endêmica em 76 países e territórios.

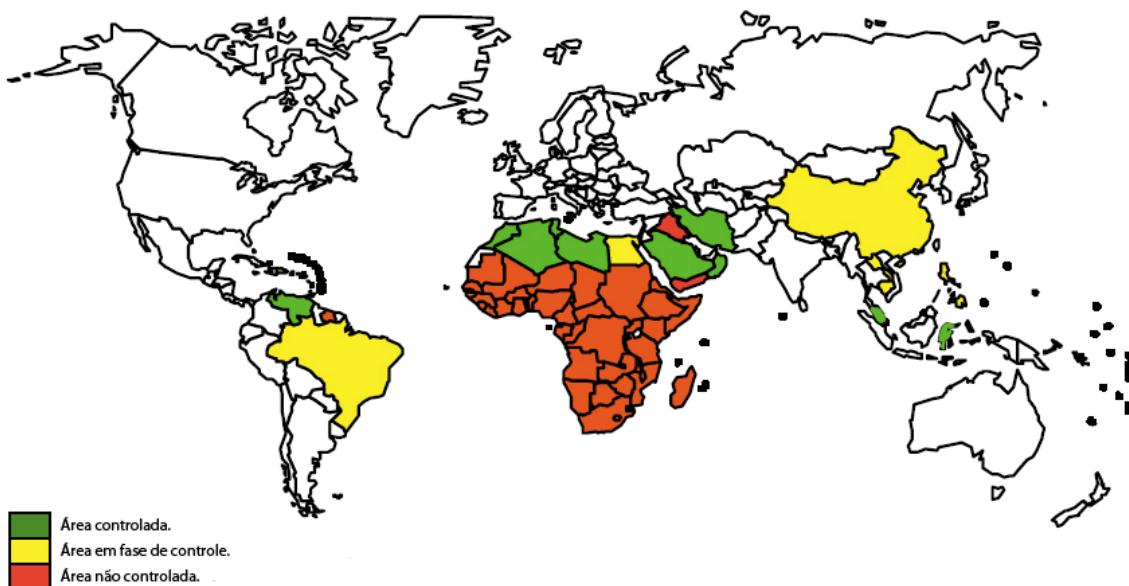


Figura 1. Estado de controle da esquistossomose no mundo (ENGELS *et al.*, 2002)

O gênero *Schistosoma* é um trematoda da família Schistosomatidae. Este gênero é formado por 4 grupos de espécies, conforme apresentado na tabela 1, sendo que apenas as espécies *S. bovis*, *S. indicum* e *S. nasale* não são parasitas em humanos (COON, 2005).

Tabela 1. Grupo de espécies de *Schistosoma* (COON, 2005)

Grupos	Espécies
<i>Schistosoma mansoni</i>	<i>S. mansoni</i>
	<i>S. japonicum</i>
<i>Schistosoma japonicum</i>	<i>S. mekongi</i>
	<i>S. malayensis</i>
	<i>S. haematobium</i>
<i>Schistosoma haematobium</i>	<i>S. intercalatum</i>
	<i>S. mattheei</i>
	<i>S. bovis</i>
<i>Schistosoma indicum</i>	<i>S. indicum</i>
	<i>S. nasale</i>

O *S. mansoni* é a espécie que apresenta a maior distribuição global (JOHNSTON, 1993). Na América do Sul, esta espécie é a causadora desta parasitose (figura 2), distribuindo-se desde países do caribe até vários estados do Brasil (GRYSEELS *et al.*, 2006).

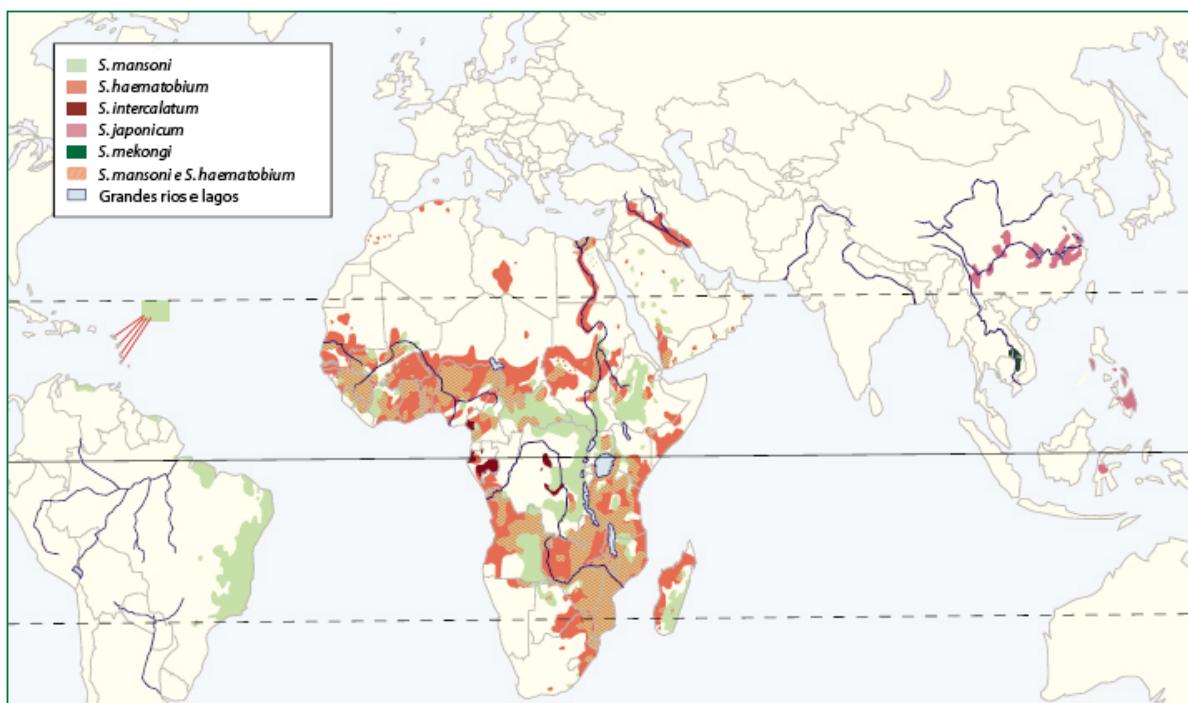


Figura 2. Distribuição global do *S. mansoni* (GRYSEELS *et al.*, 2006)

Em termos de morbidade, a esquistossomose tem papel de destaque no ranking das parasitoses de importância em saúde pública (WHO, 2005), perdendo apenas para a malária como causa de morbidade em termos de doença parasitária (CHITSULO *et al.*, 2000). A nível de Brasil, é estimado que 8 a 10 milhões de pessoas sejam afetadas por esta doença (FERRARI *et al.*, 2003). Dados oficiais mostram que no ano de 2006 foram confirmados 7.890 casos dessa doença a nível nacional (BRASIL, 2006). A nível estadual, Pernambuco possui importante prevalência da esquistossomose (BARBOSA *et al.*, 1996). Dados de 1994 mostraram que Pernambuco possuía uma área endêmica de 17.215 km², o que corresponde a 17,5% de sua área territorial, possuindo 47% de municípios afetados. (MARQUES, 1994).

A esquistossomose é uma parasitose, cujo agente etiológico possui um ciclo evolutivo do tipo heteroxênico, necessitando de dois hospedeiros para desenvolvimento de seu ciclo. O molusco do gênero *Biomphalaria* destaca-se como hospedeiro intermediário

para esta doença (REY *et al.*, 1991). No Brasil, são encontradas três espécies desse molusco que estão envolvidas com a transmissão da doença: *Biomphalaria glabrata*, *Biomphalaria straminea* e *Biomphalaria tenagophila* (PARAENSE, 1983). Em termos de distribuição, o *B. glabrata* abrange 16 estados (Alagoas, Bahia, Espírito Santo, Goiás, Maranhão, Minas Gerais, Pará, Paraíba, Paraná, Pernambuco, Piauí, Rio Grande do Norte, Rio Grande do Sul, Rio de Janeiro, São Paulo e Sergipe) e o Distrito Federal. O *B. straminea* tem distribuição conhecida mais extensa, estando presente, praticamente, em todas as bacias hidrográficas do território brasileiro. Ocorre em 23 estados (Acre, Alagoas, Amazonas, Bahia, Ceará, Espírito Santo, Goiás, Maranhão, Mato Grosso, Mato Grosso do Sul, Minas Gerais, Pará, Paraíba, Paraná, Pernambuco, Piauí, Rio Grande do Norte, Rio Grande do Sul, Rio de Janeiro, São Paulo, Santa Catarina, Sergipe e Tocantins) e no Distrito Federal. O *B. tenagophila* é amplamente encontrada no sul do país, embora possa ser detectada em menor extensão em outras regiões. Hoje, sua distribuição alcança 11 estados (Bahia, Goiás, Mato Grosso, Mato Grosso do Sul, Espírito Santo, Minas Gerais, Rio de Janeiro, São Paulo, Paraná, Rio Grande do Sul e Santa Catarina) e o Distrito Federal (FUNASA, 2002). Destas três espécies, o *B. glabrata* é o que apresenta a maior susceptibilidade à infecção pelo *S. mansoni*, devido ao seu maior tamanho, melhor adaptação ao parasita, maior sobrevida e maior numero de larvas infectantes liberadas (MALAGUEÑO *et al.*, 1994).

Já o homem, é comumente o hospedeiro definitivo da doença, embora haja, em modelo experimental, o camundongo como excelente hospedeiro. Tudo começa na água, quando as fezes humanas contaminadas com ovos do *S. mansoni* liberam o miracídio (figura 3).

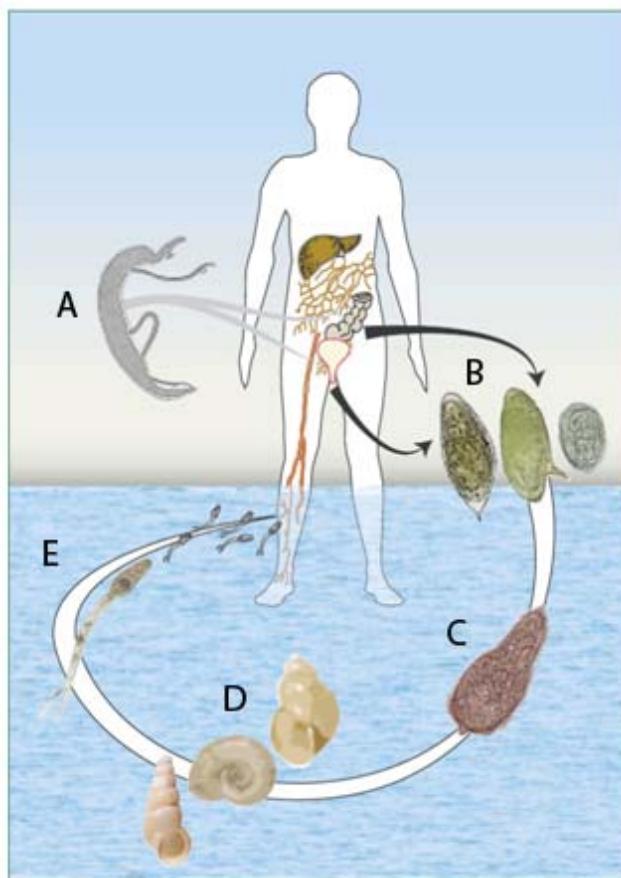


Figura 3 – Ciclo de vida do *Schistosoma mansoni* (GRYSEELS et al., 2006).

A forma larvária (figura 3 - C), o miracídio, existente no interior do ovo, em condições adequadas de luz e temperatura, liberta-se e nada ativamente ao encontro de seu hospedeiro intermediário. Neste (figura 3 - D), ocorre o desenvolvimento dos esporocistos originando as cercárias. Sob estímulos como luz, temperatura, os moluscos liberam várias cercárias (figura 3 - E), as quais nadam ativamente até encontrar seu hospedeiro definitivo. Após transpassar a pele do homem, transformam-se em esquistossômulos os quais são levados pela corrente sanguínea até o pulmão (local de alongamento). Através das veias pulmonares saem do pulmão até ganharem a grande circulação onde são conduzidos ao sistema portal-hepático (figura 3 - A). No interior dos vasos do sistema porta (figura 4), o verme evolui até tornar-se adulto. Após crescimento, ocorre o acasalamento, período que o casal migra para o sistema porta-mesentérico para oviposição.

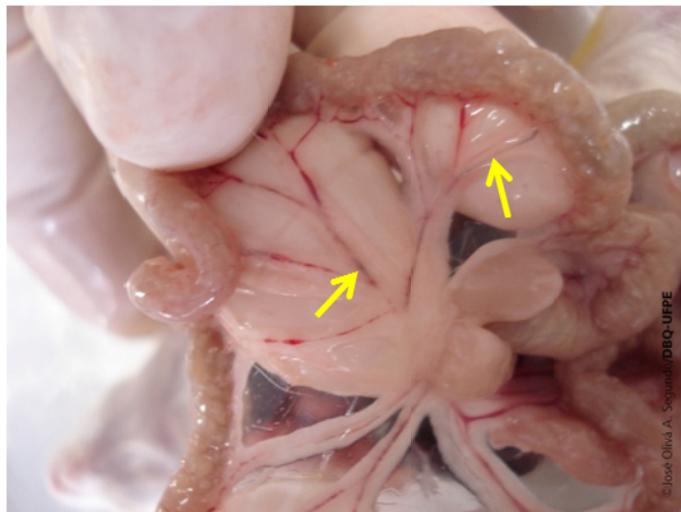


Figura 4 – Plexo mesentérico de camundongos infectados por *S. mansoni*.

A evolução e a sintomatologia desta parasitose dependem de fatores como espécie do parasito, carga parasitária, freqüência de reinfecções, idade e estado imunológico do indivídio. (KABATEREINE *et al.*, 1999). Durante o processo de oviposição, 50% dos ovos eliminados chegam à luz intestinal para serem eliminados junto com as fezes. Entretanto, os ovos que não conseguem chegar à luz intestinal são transportados via circulação, ficando retidos e depositados em órgãos como pulmão, intestino e principalmente o fígado, local das maiores alterações histopatológicas, fisiológicas e bioquímicas (COUTINHO, 1973).

Com a deposição de ovos no fígado, dá-se início à reação inflamatória visando isolar e reter os ovos maduros com a resultante formação de lesões granulomatosas peri-ovulares de localização portal, peri-portal ou parenquimatosa decorrentes da resposta imunológica aos抗ígenos liberados pelos ovos com consequente formação de fibrose hepática (COELHO, 1971). Estas áreas fibrosadas (figura 5) ao longo dos vasos hepáticos podem levar a uma fibrose perivasicular, conhecida como fibrose de Symmers, a qual desencadeará outros fenômenos como a hipertensão portal e esplenomegalia (BOGLIOLLO, 1957).



Figura 5 – Fígado contendo fibroses (pontos brancos) decorrentes de lesões granulomatosas oriundas da infecção por *S. mansoni*

A reação granulomatosa que se desenvolve ao redor dos ovos do parasito, tem como base uma hipersensibilidade do tipo celular (WARREN *et al.*, 1967), em decorrência da eliminação de抗ígenos do miracídio, chamados de SEA (*Soluble egg antigens* – Antígenos solúveis do ovo) que atravessam a casca do ovo, estimulando células como macrófagos, eosinófilos, linfócitos e citocinas como a interleucina-2 e interferon gama (HANG *et al.*, 1974; WYLER *et al.*, 1978), além também de componentes da resposta imune humoral (DESSEIN *et al.*, 1992; WEBSTER *et al.*, 1996). Contudo, as alterações hepáticas parecem não decorrer apenas do processo granulomatoso e da fibrose formada posteriormente, mas também da liberação de substâncias hepatotóxicas produzidas pelos ovos do parasito (AMIRI *et al.*, 1992).

Por conta das alterações fisiopatológicas provocadas principalmente pelo granuloma, alterações na função hepática podem ser detectadas através das dosagens da atividade de enzimas como aspartato aminotransferase (AST) e alanina aminotransferase (ALT), enzimas intracelulares que são liberadas na circulação em algumas hepatopatias, como a

esquistossomose (MANSOUR *et al.*, 1982). Estas servem para avaliar a função hepática e consequentemente o grau da lesão.

Diversos medicamentos já foram desenvolvidos e utilizados para o combate ao verme (CIOLI *et al.*, 1995). Atualmente, por não existir uma vacina contra a doença, por não existir um controle efetivo do vetor e pelas ações sanitárias ineficientes de saúde pública, o tratamento quimioterápico é a principal ferramenta no controle da esquistossomose (CAFFREY, 2007).

Dentre esses medicamentos utilizados, destaca-se o praziquantel, droga de escolha para o tratamento da esquistossomose por apresentar alta eficácia contra todas as espécies do Schistosoma, poucos efeitos adversos e administração em dose única por via oral (DOENHOFF *et al.*, 2006).

O praziquantel (PZQ) é um derivado pirazino-isoquinolona, de nome químico 2-ciclohexilcarbonil-1,2,3,6,7,11b-hexahidro-4H-pirazino-[2,1-a]isoquinolin-4-ona. Sua estrutura química é apresentada na figura 6.

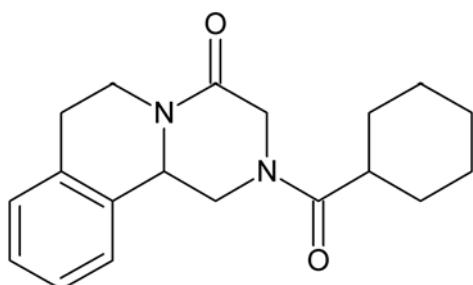


Figura 6 – Estrutura química do praziquantel.

O mecanismo de ação do PZQ ainda não está totalmente elucidado. Uma das hipóteses aceitas é que o PZQ, ao entrar em contato com verme, cause um influxo de íons

cálcio, levando a um aumento da atividade muscular com consequente contração da musculatura, levando a uma ruptura do tegumento helmíntico (PAX *et al.*, 1978; FETTERER *et al.*, 1980; BECKER *et al.*, 1980). O PZQ apresentou baixa toxicidade e mostrou-se seguro e efetivo em sua administração (CIOLI *et al.*, 2003). Porém, ele apresenta alguns efeitos secundários mínimos como dor de cabeça, desconforto abdominal (DAVIS, 1993), possuindo uma alta taxa de eficácia (DAYAN, 2003).

Outra alternativa medicamentosa pouco empregada para o controle da doença é o oxaminiquine, cuja ação no verme é inibir a síntese de seus ácidos nucléicos, diminuindo a motilidade do parasito (OLDS *et al.*, 2000). Porém, ao contrário do PZQ, essa droga mostra-se eficaz somente para a espécie *S. mansoni* (BECK *et al.*, 2001) e apresenta maior relação custo/tratamento em comparação ao PZQ que custa de 7 a 19 centavos de US\$ o comprimido de 600 mg (McFADYEN, 2006) .

Entretanto, alguns trabalhos vêm mostrando o aparecimento de cepas de *S. mansoni* resistentes ao tratamento convencional. Produzida em laboratório uma mistura de gênes de *S. mansoni* oriundas de quatro diferentes áreas geográficas, foi verificado que 93% dos vermes resistiram ao tratamento em 3 doses de 300mg/kg de PZQ (FALLON *et al.*, 1994). Em outro estudo epidemiológico, foi verificado que em uma área do Senegal, país africano cuja prevalência da doença era de 91% na população, o tratamento convencional com PZQ apresentou baixa taxa de eficácia de tratamento (18%) (STELMA *et al.*, 1995). Portanto, é necessário realizar estudos mais criteriosos para verificar outros locais de cepas resistentes ao tratamento. Embora existam outras drogas alternativas como a oxaminiquina para pronta ação, tem sido incentivados estudos visando o desenvolvimento de novos compostos

com o intuito de implementar novas drogas esquistossomicidas (UTZINGER *et al.*, 2003; XIAO *et al.*, 2007; MOREIRA *et al.*, 2007).

Nesse contexto, as imidazolidinas vêm se destacando como fármacos com boa atividade antiparasitária frente ao *S. mansoni* *in vitro* e *in vivo* (PITTA *et al.*, 2006; ALBUQUERQUE *et al.*, 2005; OLIVEIRA *et al.*, 2004).

As imidazolidinas são compostos heterocíclicos pentagonais aromáticos e apresentam em sua estrutura dois átomos de nitrogênio (figura 7) (BARREIRO *et al.*, 2001).

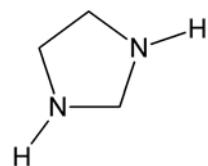


Figura 7 – Estrutura química do anel imidazolidínico.

Esses compostos, assim como seus derivados, demonstraram possuir várias atividades biológicas, dentre as principais como anticonvulsivante (THENMOZHIYAL *et al.*, 2004), antimicrobiana (LIMA *et al.*, 1992), atividade anti-hipertensiva (DYLAG *et al.*, 2004), atividade antineoplásica (ROBINSON *et al.*, 2001), além de atividade esquistossomicida já citada anteriormente.

Através de reações de síntese orgânica, é possível a adição de outros grupamentos químicos ao anel imidazolidínico. Ao adicionar dois grupamentos cetônicos nas posições 2/4 do anel (figura 8), obtém-se como produto a imidazolidina-2,4-diona ou também chamada de Hidantoína.

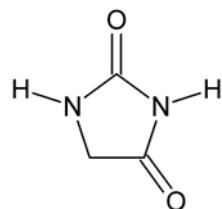


Figura 8 - Estrutura química da imidazolidina-2,4-diona

Entretanto, ao adicionar-se um grupamento tioxo na posição 4 e um grupamento cetônico na posição 2 do anel, obtemos a 4-tioxo-imidazolidin-2-ona (figura 9).

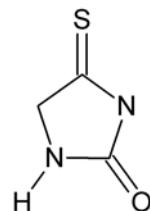


Figura 9 – Estrutura química da 4-Tioxo-imidazolidin-2-ona

O derivado 3-(4-cloro-benzil)-5-(4nitro-benzilideno)-imidazolidina-2,4-diona (LPSF-FZ4) (figura 10), cuja síntese é descrita por Lima *et al.* (2002), é obtido através da reação do brometo de benzila ou fenacila com a imidazolidina-2,4-diona, dando origem ao composto intermediário imidazolidina-2,4-diona alquilado, o qual reage com o ácido acético/acetato de sódio e benjaldeído aromático, levando a formação do derivado LPSF-FZ4, cuja algumas características químicas estão indicadas na tabela 2. Este composto mostrou-se efetivo contra vermes do *S. mansoni* *in vitro*, apresentando uma taxa de mortalidade de 100% no 9º dia de contado a uma concentração de 60 µg/mL (ALBUQUERQUE, 2002).

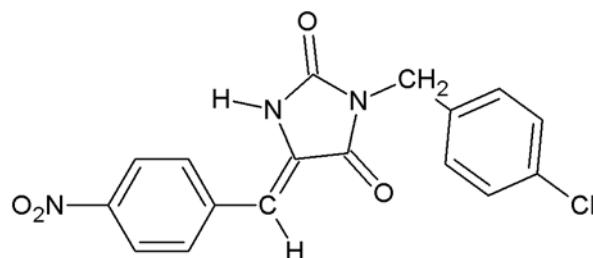


Figura 10 – Estrutura química do 3-(4-cloro-benzil)-5-(4nitro-benzilideno)-imidazolidina-2,4-diona (LPSF-FZ4).

Outro derivado, o 3-benzil-5-(4-cloro-arylazo)-4-tioxo-imidazolidin-2-ona (LPSF-PT5) (figura 11), cuja síntese é descrita por Brandão *et al.* (1997), é produzido através de três reações. A 3-benzil-imidazolidina-2,4-diona, obtida através da reação da imidazolidina-2,4-diona com o clereto de benzil reage com P_2S_5 , formando 3-benzil-4-tioxo-imidazolidin-2-ona que reage com a anilina substituída em meio ácido e na presença de nitrato de sódio, dando origem a formação de compostos arilazo-imidazolidionicos, dentre eles o LPSF-PT5, cujas algumas características químicas estão indicadas na tabela 2. Foi verificado que, através de ensaios de susceptibilidade *in vitro* frente ao *S. mansoni* (cepa BH), mostrou possuir relevante atividade esquistossomicida, causando letalidade em 100% dos vermes a partir do quarto dia após o início do tratamento a uma concentração entre 20 a 60 $\mu\text{g}/\text{mL}$ (SOARES, 2004).

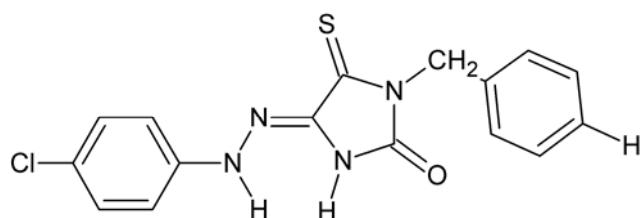


Figura 11 – Estrutura química do 3-benzil-5-(4-cloro-arylazo)-4-tioxo-imidazolidin-2-ona (LPSF-PT5)

Tabela 2 – Algumas características químicas dos compostos 3-(4-cloro-benzil)-5-(4nitro-benzilideno)-imidazolidina-2,4-diona (LPSF-FZ4) e 3-benzil-5-(4-cloro-arylazo)-4-tioxo-imidazolidin-2-oná (LPSF-PT5).

Composto	Cor	Fórmula	Massa (mols)
LPSF-FZ4	Amarelo	C ₁₇ H ₁₂ N ₃ O ₄ Cl	357,7
LPSF-PT5	Alaranjado	C ₁₆ H ₁₂ N ₄ OSCl	343,5

Os lipídios são biocompostos com diversos grupamentos químicos, cuja principal característica é sua insolubilidade em água e solubilidade em solventes orgânicos (LEHNINGER *et al.*, 2002). Os lipídios presentes no plasma, mais importante do ponto de vista fisiológico e clínico, são os ácidos graxos, fosfolipídios que são componentes das membranas biológicas, triglicerídeos (TG) que servem de armazenamento energético mais importante do organismo (presente principalmente no tecido adiposo), e colesterol (precursor dos hormônios esteróides, dos ácidos biliares, da Vitamina D além de exercer importantes funções nas membranas biológicas) (GRAUBER, 2000). Esses lipídios são transportados de um órgão para outro, utilizando o plasma sanguíneo como rede de transporte, principalmente na forma de lipoproteínas, molécula composta por lipídios e proteínas específicas (apoproteinas), tendo como exemplos principais: o quilomicron, a lipoproteína de muito baixa densidade (VLDL), a lipoproteína de baixa densidade (LDL) e a lipoproteína de alta densidade (HDL).

Estudos anteriores demonstraram que a esquistossomose promove diversas alterações no metabolismo lipídico como diminuição nos níveis de colesterol total (CT), fosfolipídios e TG plasmáticos em humanos (COUTINHO-ABATH *et al.*, 1966) (GILLETT *et al.*, 1978), em camundongos (OWEN *et al.*, 1978; VIEIRA *et al.*, 1992) e em primatas não humanos da espécie *Callithrix jacchus* (LIMA *et al.*, 1998), na fase crônica da doença. Hipóteses tais como a não produção de colesterol pelo schistosoma (MEYER *et al.*, 1970),

incorporação de partículas de LDL pelo parasita (BENNETT *et al.*, 1991) via internalização de receptores LDL (RUMJANEK *et al.*, 1983), indução da síntese de anticorpos naturais que metabolizam o colesterol através de opsonização (ALVING *et al.*, 1999) e também diminuição de atividade da enzima esterificante do colesterol na circulação, a lecitina: colesterol aciltransferase (LCAT) (LIMA *et al.*, 1998) justificam a diminuição desses lipídios.

A literatura pouco reporta sobre alterações promovidas por drogas esquistossomicidas sobre os lipídios plasmáticos. O PZQ, ao interagir com a bicamada lipídica das membranas biológicas promove uma discreta esfoliação de alguns lipídios constituintes da mesma, diminuindo discretamente o colesterol e fosfolipídios da membrana de eritrócito humano (MALHEIROS *et al.*, 2000).

Derivados imidazólicos, análogos estruturais das imidazolidinas que se diferenciam destas por possuir 4 átomos de hidrogênio a menos (figura 12), muitos desses empregados em terapias antimicrobianas, mostraram possuir atividade sobre o metabolismo lipídico (WIERZBICKI, 2003).

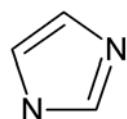


Figura 12 – Estrutura do anel imidazólico.

Dentre esses, podemos destacar o metronidazol, muitas vezes empregado para o combate de amebíase. Este fármaco promove, em humanos, diminuições significativas de 19% de colesterol LDL e de 23% de triglicerídeos, bem como discreto aumento de 10% nos níveis de colesterol HDL (SHAMKHANY *et al.*, 2003). Outros trabalhos mostraram que o cetoconazol, agente antimicótico, também possui potencial atividade hipolipidêmica, promovendo as

mesmas ações supracitadas no metabolismo lipídico, porém em percentuais menores (10% de redução de LDL colesterol e discreta ação redutora sobre triglicerídeos) (KRAEMER *et al.*, 1986; STALENHOEF *et al.*, 1997). Além disso, derivados imidazolicodinônicos promoveram alterações positivas, elevando o nível de colesterol HDL (ELOKDAH *et al.*, 2000). Em geral, os derivados imidazólicos promovem inibição das enzimas do citocromo P450, incluindo aquelas que estão envolvidas com a esterificação do colesterol, como a acil-CoA colesterol acil transferase (ACAT) (AHMED *et al.*, 1995), bem como promovem aumento de atividade de receptores LDL (BURNETT *et al.*, 1999; KEMPEN *et al.*, 1987).

Também é reportado na literatura que alguns compostos imidazolidínicos demonstraram possuir alguma atividade que promovessem alterações no metabolismo lipídico. Foi observado que a fenitoína, cujo nome químico é 5,5-difenil-imidazolidina-2,4-diona, empregada para tratamento de epilepsia, quando administrada por via oral em cavalos, promoveu elevação dos TG bem como alteração a utilização e incorporação de ácidos graxos (FLETCHER, *et al.*, 1993), elevação das concentrações plasmáticas de HDL-C em humanos (GOERDT *et al.*, 1995) e, independentemente do nível de HDL-C, reduziu o aparecimento precoce e o desenvolvimento da aterosclerose (TROCHO *et al.*, 2004). A Nitrofurantoína, de nome químico 1-(5-nitro-2-furfurilideneamino)-hidantoina, utilizada para o tratamento de infecções do trato gênito-urinário, promoveu inibição da peroxidação lipídica microssomal em ratos (DUBIN *et al.*, 1987). Também a Clonidina (n-(2,6-diclorofenil)-4,5-dihidro-1H-imidazol-2-amina), utilizada como anti-hipertensivo, causou em pacientes com hipertensão leve uma diminuição de HDL-C e de apolipoproteína A-I e apolipoproteína A-II (HOUSTON *et al.*, 1990).

Como a esquistossomose promove diminuição nos lipídios plasmáticos como CT, TG e FLT e também, devido existir poucos trabalhos mostrando o efeito de fármacos com ação esquistosomicida sobre os lipídios plasmáticos, estudos que visem observar o efeito do tratamento antiparasitário sobre o metabolismo lipídico são de extrema importância, uma vez que mudanças que promovam elevações, principalmente, nos níveis de CT e TG, estejam relacionadas com o aparecimento e desenvolvimento das doenças cardiovasculares, apontadas hoje como um grave problema de saúde pública, acarretando em milhares de óbitos em todo mundo. Drogas que demonstrem possuir ação no metabolismo lipídico, como alguns derivados imidazólicos, análogos estruturais das imidazolidinas, estão sendo apontados como potenciais coadjuvantes para terapias hipolipidêmicas. Por isso, torna-se não só importante o estudo dos derivados imidazolidínicos frente ao *S. mansoni*, mas também em relação ao metabolismo de lipídios e lipoproteínas visando elucidar novas drogas com potencial atividade hipolipidêmica.

2. OBJETIVOS

2.1. Geral

Avaliar o efeito do tratamento com os derivados imidazolidínicos 3-benzil-5-(4-cloro-
arilazo)-4-tioxo-imidazolidin-2-ona (LPSF-PT5), 3-(4-cloro-benzil)-5-(4nitro-benzilideno)-
imidazolidina-2,4-diona (LPSF-FZ4) e do Praziquantel sobre o metabolismo lipídico de
camundongos infectados e não infectados por *S. mansoni*.

2.2. Específicos

- Infectar camundongos fêmeas com o *S. mansoni* (cepa BH);
- Tratar via oral, por 5 dias consecutivos, camundongos infectados e não infectados
com os derivados imidazolidínicos LPSF-PT5 e LPSF-FZ4 e Praziquantel nas
concentrações de 50 mg/kg/dia e 100 mg/kg/dia.
- Determinar os níveis dos lipídios plasmáticos (CT, TG e FT) e atividade enzimática de
AST e ALT dos camundongos infectados e não infectados antes e após 15 dias do
tratamento com os compostos avaliados.
- Avaliar estatisticamente os resultados dos parâmetros bioquímicos obtidos entre os
grupos de estudo.

3. REFERÊNCIAS BIBLIOGRÁFICAS

AHMED, S. et al. Synthesis and biological evaluation of imidazole based compounds as cytochrome P-450 inhibitors. *Drug design and discovery*, v. 13, p.27-41, 1995.

ALBUQUERQUE, M.C.P.A. Novas imidazolidinas potencialmente ativas no combate à esquistossomose: síntese e avaliação da atividade no Schistosoma mansoni (cepa BH). 2002. Tese (Doutorado) - Programa de Pós-Graduação em Ciencias Biológicas, Universidade Federal de Pernambuco, Recife, 2002.

ALBUQUERQUE, M.C.P.A. et al. Synthesis and schistosomicidal activity of new substituted thioxo-imidazoline compounds. *Die Pharmazie*, v.60, n.1, p. 13-17, 2005.

ALVING, C. R.; WASSEF, N. M. Naturally occurring antibodies to cholesterol: a new theory of LDL cholesterol metabolism. *Immunology Today*, v. 20, p. 362-366, 1999.

AMIRI, P. et al. Tumour necrosis factor alpha restores granulomas and induces parasite egg-laying in schistosome-infected SCID mice. *Nature*, v. 356, n.6370, pp. 604-607, 1992.

BARBOSA, C.S.; SILVA, C.B.; BARBOSA, F.S. Esquistossomose: reprodução e expansão da endemia no estado de Pernambuco no Brasil. *Revista de Saúde Pública*, v.30, n.6, p. 609-616, 1996.

BARREIRO, E.J.; FRAGA, C.A.M. *Química Medicinal: As bases moleculares da ação dos Fármacos*. Porto Alegre : Artmed Editora, 2001, 243 p.

BECK, L. et al. Replacing oxamniquine by praziquantel against Schistosoma mansoni infection in a rural community from the sugar-cane zone of Northeast Brazil: an epidemiological follow-up. *Memórias do Instituto Oswaldo Cruz*, v.96, p. 165-167, 2001.

BECKER, B. et al. Light and electron microscopic studies on the effect of praziquantel on Schistosoma mansoni, Dicrocoelium dendriticum, and Fasciola hepatica (Trematoda) in vitro. *Zeitschrift für Parasitenkunde*, v.63, p. 113-128, 1980.

BENNETT, M.W.; CAULDFILD, J.P. Specific binding of human low-density-lipoprotein to the surface of schistosomula of Schistosoma mansoni and ingestion by the parasite. *American Journal of Pathology*, v.131, p. 1173-1182, 1991.

- BLANCHARD, TOM J. Schistosomiasis. *Travel Medicine and Infectious Disease*, v.2, 2004.
- BOGLIOLO, L. Anatomical picture of liver in hepatosplenic schistosomiasis mansoni. *Ann Trop Med Parasitol*, v. 51, p. 1-14, 1957.
- BRANDÃO, S.S.F. et al. Synthèse et structure des arylazo-imidazolidines et arylidènethiazolidines substituées. *Ann Pharmaceutiques Françaises* 1997, v.55, n.5, p.206-211, 1997.
- BRASIL, Ministério da Saúde. Esquistossomose - Casos confirmados notificados no Sistema de Informação de Agravos de Notificação – Sinan em 2006. Disponível em <<http://dtr2004.saude.gov.br/sinanweb/novo/>>. Acesso em: 30 jan. 2008.
- BURNETT, J.R.; WILCOX, L.J.; HUFF, M.W. Acyl coenzyme A: cholesterol acyltransferase inhibition and hepatic apolipoprotein B secretion. *Clinica chimica acta*, v.286, p. 231-242, 1999.
- CAFFREY, C.R. Chemotherapy of schistosomiasis: present and future. *Chemical Biology*, v. 11, p.433-439, 2007.
- CHITSULO, L. et al. The global status of schistosomiasis and its control. *Acta Tropica*, v. 77, p. 41-51, 2000.
- CIOLI, D.; PICA-MATTOCCIA, L. Praziquantel. *Parasitology Research*, v.90, n.1, p. 3-9, 2003.
- CIOLI, D.; PICA-MATTOCCIA, L.; ARCHER, S. Antischistosomal drugs: Past, present ... and future? *Pharmacology & Therapeutics*, v.68, n.1, p. 35-85, 1995.
- COELHO, R.B. *Anatomia Patológica das Afecções Hepáticas*. Recife : Imprensa Universitária, Universidade Federal de Pernambuco, 1971.
- COON, DAVID R. Schistosomiasis: Overview of the history, biology, clinicopathology, and laboratory diagnosis. *Clinical Microbiology Newsletter*, v. 27, n.21, 2005.
- COUTINHO, A. Fígado e Esquistossomose. *Jornal Brasileiro de Medicina*, p. 23-41, 1973.
- COUTINHO-ABATH, E.; BARBOSA, J.M.; AMARAL, J.A. Alterações bioquímicas na esquistossomose mansônica humana, como especial referência ao metabolismo lipídico. *Jornal Brasileiro de Medicina*, v. 11, n.2, p.157-168, 1996.
- DAVIS, A. Antischistosomal drugs and clinical practice. In: P. JORDAN; G. WEBBE; R.F. STURROCK. Wallingford : CAB International, 1993. p. 367-404.

- DAYAN, A.D. Albendazole, mebendazole and praziquantel: review of non-clinical toxicity and pharmacokinetics. *Acta Tropica*, v. 86, p.141-159, 2003.
- DESSEIN, A.J. *et al.* Environmental, genetic and immunological factors in human resistance to Schistosoma mansoni. *Immunol Invest*, v.21, n.5, p.423-453, 1992.
- DOENHOFF, M.J.; PICA-MATTOCCIA, L. Praziquantel for the treatmentof schistosomiasis: its use for control in areas with endemic disease and prospects for drug resistance. *Expert review of anti-infective therapy*, v.4, p.199-210, 2006.
- DUBIN, M. *et al.* Nitrofuran inhibition of microsomal lipid peroxidation. *Federation of European Biochemical Societies letters*, v. 220, n.1, p. 197-200, 1987.
- DYLAG, T. *et al.* Synthesis and evaluation of in vivo activity of diphenylhydantoin basic derivatives. *European Journal of Medicinal Chemistr*, v.39, p. 1013-1027,2004.
- ELOKDAH, H. *et al.* Effects of 2- (substituted-sulfanyl)-3,5-dihydro-imidazole-4-one and 2-(substituted-sulfanyl)-1H-imidazole-4,5-dione derivatives on serum HDL-cholesterol. *Bioorganic & medicinal chemistry letters*, v.10, p.1791-1794, 2000.
- ENGELS, D. *et al.* The global epidemiological situation of schistosomiasis and new approaches to control and research. *Acta Tropica*, v.82, p. 139-146, 2002.
- FALLON, P.G.; DOENHOFF, M.J. Drug-resistant schistosomiasis: resistance to praziquantel and oxamniquine induced in Schistosomu munsoni in mice is drug specific. *American Journal of Tropical Medicine and Hygiene*, v. 51, p. 83-88, 1994.
- FERRARI, M.L.A.; COELHO, P.M.Z. Efficacy of oxamniquine and praziquantel in the treatment of Schistosoma mansoni infection: a contrelled trial. *Bulletin of the World Health Organization*, v.81, n.3, 2003.
- FETTERER, RH.; PAX, RA.; J.L., BENNETT. Praziquantel, potassium and 2,4-dinitrophenol: analysis of their action on the musculature of Schistosoma mansoni. *European Journal of Pharmacology*, v.64, p. 31-38, 1980.
- FLETCHER, E.J.; KIRSTEN, E.; BEECH, J. Increases specific triacylglycerol fatty esters in skeletal muscle from horses with hyperkalemic periodic paralysis. *Biochimica et Biophysica Acta (BBA) - Lipids and Lipid Metabolism*, v.1168, n.3, p.292-298, 1993.

FUNASA. *Guia de Vigilância Epidemiológica*. 5.ed. Brasília : s.n., 2002. p. 282-286. Vol. I.

GILLETT, M. P. T.; CARVALHO, V.C. *Schistosoma mansoni*: A comparative study of plasma and erythrocyte lipid alterations in the experimentally infected mouse and in selected human patients. *Experimental Parasitology*, v. 44, p.173-180, 1978.

GOERDT, C.; KEITH, M.; RUBINS, H.B. Effects of phenytoin on plasma high-density lipoprotein cholesterol levels in men with low levels of high-density lipoprotein cholesterol. *Journal of Clinical Pharmacology*, v.35, p.767-775, 1995.

GRAUBER, D. S. Estimativa dos Lípides Totais no soro baseada nas Determinações de Colesterol e/ou Triglicérides. *Revista Brasileira de Análises Clínicas*, v.32, n.2, p.97-99, 2000.

GRYSEELS, B. et al. Human schistosomiasis. *The Lancet*, ed. 9541, v. 368, p. 1106-1118, 2006.

HANG, L.M.; WARREN, K.S ; BOROS, D.L. *Schistosoma mansoni*: antigenic secretions and the etiology of egg granulomas in mice. *Experimental Parasitology*, v. 35, n.2, p.288-298, 1974.

HOUSTON, M.C. et al. The effects of clonidine hydrochloride versus atenolol monotherapy on serum lipids, lipid subfractions, and apolipoproteins in mild hypertension. *American Heart Journal*, v.120, n. 1, p. 172-179, 1990.

JOHNSTON, D.A. Opening the con of worms: molecular analysis of Schistosomiasis mansoni population. *Parasitology today*, v.9, p.286-291, 1993.

KABATEREINE, N.B. et al. Adult resistance to schistosomiasis mansoni: age-dependence of reinfection remains constant in communities with diverse exposure patterns. *Parasitology*, v.111, p.101-105, 1999.

KEMPEN, H.J. et al. Effect of ketoconazole on cholesterol synthesis and on HMG-CoA reductase and LDL-receptor activities in Hep G2 cells. *Biochemical pharmacology*, v.36, p. 1245-1249, 1987.

KRAEMER, F.B.; SPILMAN, S.D. Effects of ketoconazole on cholesterol synthesis. *Journal of pharmacology and experimental therapeutics*, v.238, p.905-911, 1986.

LEHNINGER, A. L.; NELSON, D.L. COX, M.M. *Lehninger Principles of Biochemistry*. 4 ed., p. 343, 2002.

LIMA, M.C.A, et al. Synthèse et activité antimicrobienne des chlorobenzyl benzylidène imidazolidinediones at thiazolidinediones substituées. *Pharmazie*, v. 4, n.3, p. 182-184, 1992.

LIMA, V.L.M. et al. An evaluation of Callithrix jacchus (saguí) as an experimental model for the dyslipoproteinemia of human Schistosomiasis mansoni. *Biochimica Biophysica Acta*, v. 1393, n.2, p.235-243, 1998.

MALAGENÓ, E.; SANTANA, J.V. Etiologia. In: *Esquistossomose Mansônica*, J. MALTA. Recife : Editora Universitária, 1994, p.26-46.

MALHEIROS, S.V.P. et al. Membrane effects of trifluoperazine, dibucaine and praziquantel on human erythrocytes. *Chemico-Biological Interactions*, v.126, p. 79-95, 2000.

MANSOUR, M.M. et al. Serum enzyme tests in hepatosplenic schistosomiasis. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, v. 76, n.1, p.109-111, 1982.

MARQUES, R.J. *Esquistossomose mansônica - Súmula histórica do que tem sido estudado sobre o assunto em Pernambuco*. 1ed. Recife : Editora Universitária, 1994. p. 17-22.

McFADYEN, J. International Drug Price Indicator Guide. *Management Science for Health*. 2006.

MEYER, F.; MEYER, H.; BUEDING, E. Lipid metabolism in the parasitic and free-living flatworms, *Schistosoma mansoni* and *Dugesta dorotocephalia*. *Biochimica et Biophysica Acta*, n.210, p. 257-266, 1970.

MOREIRA, L.S. et al. A study of the activity of 2-(alkylamino)-1-phenyl-1-ethanethiosulfuric acids against infection by *Schistosoma mansoni* in a murine model. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, v. 101, p.385-390, 2007.

OLDS, G.R.; DASARATHY, S. Schistosomiasis. *Current Treatment Options in Infectious Diseases*, v.2, p. 88-99, 2000.

OLIVEIRA, S.M. et al. 2004. A resposta do *Schistosoma mansoni* mantido in vitro frente a derivados imidazolidinônicos. *Acta Farmaceutica Bonaerense*, v.23, n.3, p.243-248, 2004.

OWEN, J.S. et al. Effect of experimental schistosomiasis mansoni on plasma and erythrocyte lipids and on plasma lecithin: cholesterol acyltransferase in the mouse. *Revista Brasileira de Biologia*, v.38, n.4, p.913-918, 1978.

PAX, R, J.L.; BENNETT e FETTERER, R. A benzodiazepine derivative and praziquantel: effects on musculature of *Schistosoma mansoni* and *Schistosoma japonicum*. *Naunyn Schmiedebergs Arch Pharmacol*, n.304, p.309-315, 1978.

PARAENSE, W.L. Distribuição dos caramujos no Brasil. In: REIS, F.A.; FARIA, I.; KATZ , N. Modernos Conhecimentos sobre Esquistossomose Mansônica. Suplemento dos Anais 1983/84, v. 14, Belo Horizonte: Academia Mineira de Medicina, 1983, p. 117-128.

PITTA, M.G.R. *et al.* New imidazolidinic bioisosters: potential candidates for antischistosomal drugs. *Memórias do Instituto Oswaldo Cruz*, v. 101, n.5, p.313-316, 2006.

REY, L.; PESSOA, S.B. Contribuição aos estudos dos focos de *Australorbis glabratus* em Sergipe. *Rev. Clim*, v.29, p.85-108, 1991.

ROBINSON, P.R. *et al.* Design and Synthesis of 2-Oxo-imidazolidine-4-carboxylic Acid Hydroxyamides as Potent Matrix Metalloproteinase-13Inhibitors. *Bioorganic & Medicinal Chemistry Letters*, v. 11, p. 1211-1213, 2001.

RUMJANEK, F. D.; MCLAREN, D. J.; SMITHERS, S. R. Serum-induced expression of a surface protein in schistosomula of *Schistosoma mansoni* - a possible receptor for lipid uptake. *Molecular and Biochemical Parasitology*, v.9, p.337-350, 1983.

SHAMKHANY, K.; AZARPIRA, M.; AKBAR, M.H. An open label crossover trial of effects of metronidazol on hyperlipidaemia. *International journal of cardiology*, v.90, p. 145-146, 2003.

SOARES, A.L.M. Síntese e Avaliação da Atividade Biológica de Novos Derivados Arilazo-Imidazolidínicos e Arilideno-Tiazolidínicos frente a Vermes Adultos de *Schistosoma mansoni* (Cepa BH). 2004. Dissertação (Mestrado) - Pós-graduação em Biotecnologia de Produtos Ativos, Universidade Federal de Pernambuco, Recife, 2004.

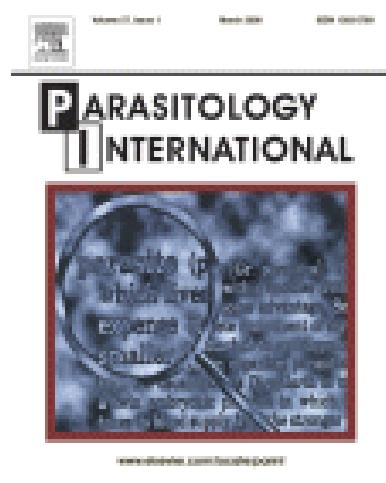
STALENHOEFF, A.F.; STUYT, P.M.; DE GRAAF, J. Effects of ketoconazole on plasma lipids and lipoprotein(a) in familial hypercholesterolaemia compared with simvastatin. *Netherlands journal of medicine*, v.51, p.10-15, 1997.

STELMA, F.F. *et al.* Low efficacy and tolerance of praziquantel in a new, intense focus of *Schistosomu mansoni* infection. *American Journal of Tropical Medicine and Hygiene*, v.53, n.2, p.167-170, 1995.

- THENMOZHIYAL, J.C.; WONG, P.T.H. ; ECHUI, W.K. Anticonvulsant activity of phenylmethylenehydantoins: a structure-activity relationship. *Journal of Medicinal Chemistry*, v. 47, p. 1527-1535, 2004.
- TROCHO, C. *et al.* Phenytoin treatment reduces atherosclerosis in mice through mechanisms independent of plasma HDL-cholesterol concentration. *Atherosclerosis*, v. 174, p. 275-285, 2004.
- UTZINGER, J. *et al.* Combination chemotherapy of schistosomiasis in laboratory studies and clinical trials. *Antimicrobial Agents and Chemotherapy*, v. 47, p. 1487-1495, 2003.
- VIEIRA, E.M. *et al.* Mouse Plasma Phospholipid Metabolism: Contributions of Lecithin: Cholesterol Acyltransferase, Phospholipase A and Lysophospholipase Activities. *Arq. Biol. Tecol.*, v. 35, n.3, p. 517-529, 1992.
- WARREN, K.S.; DOMINGO, E.O.; COWAN, R.B. Granuloma formation around schistosome eggs as a manifestation of delayed hypersensitivity. *American Journal of Pathology*, v. 51, n. 5, p. 735-756, 1967.
- WEBSTER, M. *et al.* Human immunoglobulin E responses to a recombinant 22.6- kilodalton antigen from *Schistosoma mansoni* adult worms are associated with low intensities of reinfection after treatment. *Infect Immun*, v.64, n.10, p. 4042-4046, 1996.
- WHO. Report of the Scientific Working Group on Schistosomiasis. *Programme for Research & Training in Tropical Diseases (TDR)*, 2005.
- WIERZBICKI, A.S. Imidazole derivatives as cholesterol-lowering agents. *International Journal of Cardiology*, v. 90, p. 145-146, 1993.
- WYLER, D.J.; WAHL, S.M ; WAHL, L.M. Hepatic fibrosis in schistosomiasis: egg granulomas secrete fibroblast stimulating factor in vitro. *Science*, ed. 4366, v.202, p. 438-440, 1978.
- XIAO, S.H *et al.* In vitro and in vivo activities of synthetic trioxolanes against major human schistosome . *Antimicrobial Agents and Chemotherapy*, v. 51, p. 1440-1445, 2007.

**4. CAPÍTULO I - EFFECT OF TREATMENT WITH NEW IMIDAZOLIDINE DERIVATIVES
AND PRAZIQUANTEL ON PLASMA LIPID OF MICE INFECTED BY *SCHISTOSOMA
MANSONI***

Este artigo será submetido à revista internacional “Parasitology International”
Fator de impacto: 1,500



Effect of treatment with new imidazolidine derivatives and praziquantel on plasma lipid of mice infected by *Schistosoma mansoni*

J. O. Apolinário Segundo^a, M. C. A. Lima^b, M. C. P. Albuquerque^c, I. R. Pitta^b, S. L. Galdino^b, V. L. M. Lima^{a*}

^a Laboratório de Química e Metabolismo de Lipídios e Lipoproteínas, Departamento de Bioquímica, Universidade Federal de Pernambuco, Recife, Pernambuco, Brazil

^b Laboratório de Planejamento e Síntese Farmacêutica, Departamento de Antibióticos, Universidade Federal de Pernambuco, Recife, Pernambuco, Brazil

^c Departamento de Medicina Tropical, Universidade Federal de Pernambuco, Recife, Pernambuco, Brazil

* Corresponding author:

Vera Lúcia de Menezes Lima. Avenida Professor Moraes Rêgo, S/N, Cidade Universitária, Recife, Pernambuco, Brazil. CEP: 50670-420. Telefone: +558121268540. E-mail: vlm@ufpe.br

Abbreviations: AST, aspartate aminotransferase; ALT, alanine aminotransferase; CMC, Carboxymethylcellulose; TC, total cholesterol; TG, triglycerides; PZQ, praziquantel.

4.1. Abstract

The Schistosomiasis mansoni is responsible for an elevated mortality in world. The parasitosis promotes modifications in lipid metabolism such as low levels of plasma total cholesterol (TC) triglycerides (TG) and total phospholipids (TPL) and it is not reported alterations on plasma lipids induced by anti-schistosomal drugs. Imidazolic compounds are analogous structural of imidazolidines and presents hypolipidemic activity. In this work, it was evaluated the effect on plasma lipids of oral treatment for 5 days with new imidazolidine compounds 3-benzyl-5-(4-chloro-arylazo)-4-thioxo-imidazolidin-2-one (LPSF-PT5), 3-(4-chloro-benzyl)-5-(4nitro-benzylidene)-imidazolidine-2,4-dione (LPSF-FZ4) and Praziquantel (PZQ) at concentration of 50 mg/kg/day and 100 mg/kg/day with *Schistosoma mansoni* mice and also uninfected mice. PZQ and LPSF-PT5 significantly decrease by 24.59% and by 18.60% TC levels, respectively as well by 31.60%, 31.50% TG levels, respectively from plasma of infected mice treated only at 100 mg/kg/day. In uninfected groups it was not observed a significant reduction on lipids evaluated. LPSF-FZ4 at concentration of 50 mg/kg/day and 100 mg/kg/day promoted significant reductions about 44.80% and about 40.30%, respectively only on plasma TG levels from *S. mansoni* mice. Nevertheless, in both concentrations, it was observed that LPSF-FZ4 significantly decreased plasma TG level by 26.7% and by 21.7% from uninfected animals. The results suggest that similar to PZQ treatment with 50mg/kg/day and 100mg/kg/day, LPSF-PT5 does not affect the plasma CT, TG and TPL when treated for 5 days. On the other hand, LPSF-FZ4 in both evaluated doses showed to alter TG levels, candidate as a potential hypotriglyceridemic drug.

Keywords: schistosomiasis mansoni, total cholesterol, hipotrygliceridemic activity, praziquantel, imidazolidines derivatives

4.2. Introduction

Schistosomiasis mansoni is the second human parasitosis more prevalent in the world [1] which affect about 200 million of people [2]. In Brazil, it is estimated about 8 to 10 million of individual is affected by *S. mansoni* [3], the more difunded specie [4]. By depositing eggs of parasites in the liver, place of the larger histopathologic, physiological and biochemistry alterations, the worm leads to formation of granulomatous lesions [5], promoting alterations in the hepatic function, detected by intracellular enzyme activity assay as aspartate aminotransferase (AST) and alanine aminotransferase (ALT) [6]. The disease induces alterations on lipid metabolism such as decrease in plasma total cholesterol (TC) and triglycerides (TG) [7], low levels of plasma total phospholipids (TPL) in humans [8], mice [9] [10] and on non-human primates of specie *Callithrix jacchus* (sagui) [11].

Due to absence of vaccines against worms, chemotherapy is the mainly alternative employed for the treatment [12]. Praziquantel (PZQ) is the principal drug active against all schistosome species by single oral dose administration [13]. Nevertheless, some *S. mansoni* strains are resistant to the conventional treatment [14] [15], encouraging the development of new anti-schistosomal drugs [16] [17]. Imidazolidines are aromatic pentagonal heterocycle compounds [18] which have several biological activities such as anticonvulsivant [19], antihypertensive [20], anticancerous [21] and anti-schistosomal activity for *S. mansoni* [22] [23] [24].

It is not reported alterations on plasma lipids induced by anti-schistosomal drugs. Metronidazol and other imidazolic derivatives which are analogous structural of imidazolidines, used in antimicrobiotic therapies, present activity hypolipidemic [25] in humans [26]. Some common used imidazolidines compounds has activity on lipid and lipoproteins metabolisms such as fenitoin [27] [28] [29], nitrofurantoin [30] and clonidine [31].

This work shows the effects of treatment with the new imidazolidine derivatives 3-benzyl-5-(4-chloro-arylazo)-4-thioxo-imidazolidin-2-one (LPSF-PT5) and 3-(4-chloro-benzyl)-

5-(4nitro-benzylidene)-imidazolidine-2,4-dione (LPSF-FZ4) as well as with Praziquantel on lipid metabolism of schistosomiasis mansoni infected mice.

4.3. Materials and Methods

4.3.1. Drugs

The compounds 3-(4-chloro-benzyl)-5-(4nitro-benzylidene)-imidazolidine-2,4-dione (LPSF-FZ4) and 3-benzyl-5-(4-chloro-arylazo)-4-thioxo-imidazolidin-2-one (LPSF-PT5) (Figure 1) were synthesized at the LPSF (Laboratório de Planejamento e Síntese de Fármacos, Departamento de Antibióticos, UFPE, Recife, Brazil). The synthesis and physical chemistry characters from compounds has been described by Lima et al.(1992) [32] and Brandão et al. (1997) [33], respectively. Praziquantel (PZQ) from Sigma© was utilized for this work.

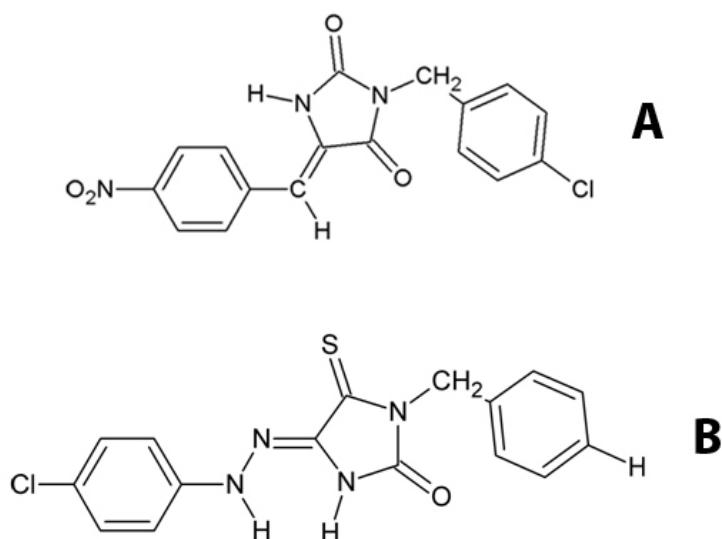


Figure 1

4.3.2. Animals

Female Swiss albino mice, 25 days old were kept under standard environmental conditions (22 ± 2 °C; 12/12 h light/dark cycle). The animals were fed with a standard diet (Labina Purina®, Brazil) and water *ad libitum*. The experimental procedure was approved by the Local Committee for Animal Ethics (Protocol 137/2003-CEEA/CCB).

4.3.3. Animal infection

The infection was carried out by tail immersion technique according to the method of Duvall and Dewitt (1967) [34] in mice of 25 days old, through the exposure to 80±10 cercariae/mouse of a BH (BH – Belo Horizonte, Brazil) strain of *S. mansoni*. The infected mice were kept under standard conditions for 49 days before start treatment.

4.3.4. Animal groups and treatment

Groups ($n = 8$) of infected and uninfected mice, by experimental antiparasitary treatment were orally treated during 5 days with 50mg/kg/day and 100mg/kg/day LPSF-PT5, LPSF-FZ4 and PZQ solubilized in 0.5% (w/v) carboxymethylcellulose (CMC) (from Sigma©). As control, group of infected and uninfected mice received oral CMC for the same period. Therefore, animals were kept under standard conditions for 15 days after treatment, sacrificed on the next day. In uninfected groups, treatment started at 75 days old.

4.3.5. Blood collection

Fasting blood collection was collected before treatment and after 15 days one. Blood (250µl) was taken from the retro-orbital sinus with a capillary utilizing brief ether anesthesia, anticoagulated with EDTA (1mg/ml) and kept in ice before using. All blood samples were centrifuged at 2,500 $\times g$ at 4°C for 18 minutes to separate erythrocytes and plasma which was used for the experiments.

4.3.6. Biochemical parameters analysis

Plasma TC and TG levels were measured by enzymatic colorimetric methods (Labtest Diagnostica, Brazil/SA). Plasma lipid extract was obtained according manufacturer instructions kit for measure TPL (Labtest Diagnostica, Brazil/SA). AST and ALT activities were determined by enzymatic colorimetric methods (Labtest Diagnostica, Brazil/SA).

4.3.7. Statistical analysis

Values are expressed as means \pm S.E.M. For the comparison between infected and uninfected mice groups before treatment it was used unpaired t test. For investigation of effect of treatment, before and after 15 days, in infected and uninfected mice groups for each concentration used was utilized Two-way ANOVA. It was used 4.0 version of GraphPad Prism computer software for analysis. Significant differences are suitable for values of probability $p < 0.05$, $p < 0.01$ and $p < 0.001$.

4.4. Results

4.4.1. Plasma Lipids

50 days after *S. mansoni* infection, mice presented significant ($p < 0.001$) low plasma TC levels when compared to uninfected mice (table 1) before treatment. Table 2 shows the comparison between plasma TC levels before and after 15 days of treatment with analyzed compounds. The results shown that in infected mice, the treatment with 100mg/kg/day of PZQ and LPSF-PT5 induced a significant decrease in plasma TC levels by 24.59% ($p < 0.001$) and by 18.60% ($p < 0.01$), respectively. All the three drugs did not significantly decrease TC levels plasma of uninfected mice.

Schistosomiasis mansonic induced significant ($p < 0.001$) decrease on plasma TG levels when compared to uninfected animals before treatment (table 1). In infected mice, treatment with 100mg/kg/day of PZQ, LPSF-PT5 and LPSF-FZ4 promoted significant decrease about 31.60% ($p < 0.01$), 31.50% ($p < 0.01$) and 40.30% ($p < 0.001$), respectively as well about 44.80% ($p < 0.001$) when treated with 50mg/kg/day of LPSF-FZ4 on plasma TG levels (table 3). However, only 50 mg/kg/day and 100mg/kg/day LPSF-FZ4 promoted significantly decreasing on plasma TG levels by 26.7% ($p < 0.05$) and by 21.7% ($p < 0.05$), respectively in uninfected mice.

Furthermore, in this work, the treatment with all evaluated compounds at dose of 50mg/kg/day and 100mg/kg/day did not promote alterations on plasma TPL levels. The infected mice TFL levels were slightly decreased in comparison to uninfected mice before treatment (table 1).

Groups which received CMC not significantly decrease TC, TG and TPL plasma levels.

4.4.2. Enzyme activities

S. mansoni infected mice presented significant ($p < 0.001$) rise AST and ALT activities in relation to uninfected mice before treatment (table 1). The infected and uninfected groups treated with 100 mg/kg/day PZQ demonstrated a significant increases by 40.92% ($p < 0.01$) and by 37.80% ($p < 0.01$) respectively in AST enzyme activity (table 4). Treatment with imidazolidines derivatives LPSF-PT5 and LPSF-FZ4 as well CMC not promoted significantly modifications in the AST enzyme activity.

Treatment with all evaluated compounds in both concentrations did not promote significantly alterations on ALT enzyme activity (table 5).

4.5. Discussion

4.5.1. Biochemical alterations

According to various factors such as parasite species, parasitic load, frequency of reinfections, age and individual immunological state [35], the inflammatory reaction caused by egg deposit in liver results in the formation of fibrosis which throughout the vase can lead to the perivascular fibrosis known as fibrosis of the Symmers which will take another phenomenons like portal hypertension and esplenomegaly [36]. In this work, *S. mansoni* infected mice presented elevated AST and ALT enzyme activities, indicating hepatic dysfunction. This result is in agreement with hepatic parameters found in plasma of mice with *S. mansoni* [37].

It has been reported that infection by *S. mansoni* alters plasma lipid levels. The low levels of plasma TC found in present work has several possibilities such as not production of cholesterol by schistosomes [38], incorporation of low density lipoprotein (LDL) by the parasite [39] via inducible LDL receptors [40], induction of synthesis of natural antibodies that metabolize cholesterol by opsonization [41] as also reduction of the enzyme activity of lecithin: cholesterol acyltransferase (LCAT) [42] which catalyse the esterification of cholesterol on plasma. Carbohydrate metabolism is affected by *S. mansoni* infection by decreased assimilation of glucose in humans [43] such as hypoglicemy in mice [44] which causes greater mobilization of TG stored, leading to decreases ones. This justify the low plasma TG level found. Increased plasma lipid levels such as TC and TG are associated to

development of heart diseases. However, effects produced by schistosomiasis on lipid metabolism promote a significant reduction in development of atherosclerotic lesions from *S. mansoni* infected mice [45]. Therefore, it is necessary to evaluate the effects produced by conventional and new anti-schistosomal drugs on plasma lipids.

4.5.2. Effects produced by treatment

Still today, PZQ is the main clinical alternative against schistosomiasis. This drug interacts with lipid layer of biological membranes and promotes a discrete exfoliation of some lipids constituent of erythrocyte layer human, decreasing slightly the cholesterol and phospholipids [46]. In this work, treatment with 100mg/kg/day PZQ promoted significant decrease on plasma TC and TG from *S. mansoni* infected mice. However, the sum of TC reduction percentual from infected control mice, plus slight TC reduction percentual from uninfected group treated with 100mg/kg/day PZQ is practically equal to TC reduction percentual found. Similarly, the significant TG reduction percentual found in infected animals treated with 100mg/kg/day PZQ is justified by same reasoning. Therefore, 100mg/kg/day PZQ treatment for 5 days did not induce alterations on plasma lipids analyzed. But, maybe if the 500 mg/kg PZQ treatment was carried out for a more prolonged time, for example 10 days, plasma TC and TG reduction percentuals obtained were bigger because PZQ effect is not dependent on the maximum drug concentration to which site is exposed, but rather on the length of time during which site is exposed to a threshold drug concentration [47]. Even if AST activity was significantly increased (table 4), 100 mg/kg/day PZQ treatment used in this work was not demonstrated to alter hepatic function. Probably there was discrete toxic action of PZQ in other organs whose AST concentrations are higher than in liver, such as heart, esqueletic muscle, spleen and kidney. Therefore, it is necessary specific studies to evaluate this possibility.

Some imidazolidine derivatives act by inhibiting P450 enzymes, including those involved with cholesterol esterification such as acyl-CoA cholesterol acyl transferase (ACAT) [48] and by enhancing LDL receptor activity [49] [50]. The mechanism of action of imidazolidines is not well established. An example of the hypothesis placed for PZQ, results with 100mg/kg/day LPSF-PT5 points for a probable hypocholesterolemic and hypotriglyceridemic actions, since that treatment be prolonged because treatment with 100 mg/kg/day 2-substituted sulfanyl-

3,5-dihydro-imidazole-4-one derivatives for 8 days promoted significant changes in serum lipids levels from rats [51]. Furthermore studies are requested for investigate this hypotheses. Nevertheless, according to our results, LPSF-FZ4 did not reduce TC levels from plasma of uninfected animals in none of the doses evaluated, although was observed a slight TC reduction on plasma from infected group treated with 50 mg/kg/day and 100 mg/kg/day LPSF-FZ4. These decreases obted is due for *S. mansoni* infection (table 2). On the other hand, results with 50 mg/kg/day LPSF-FZ4 showed a significant diminution on plasma TG from infected and uninfected mice after 15 of treatment. Significant plasma TG reductions obted with 50 mg/kg/day and 100mg/kg/day LPSF-FZ4 (table 3) from infected mice probably is not only due to infection by *S. mansoni* but too by effect of treatment. This indicates a potential hypotriglyceridemic action, requesting further confirmation. Treatment with compounds LPSF-PT5 and LPSF-FZ4 in both concentrations did not promote significantly alterations on AST and ALT enzyme activities. This found shows that treatment does not modify hepatic function in both groups of infected and uninfected mice even observed a slight reduction of AST and ALT activities at 50 mg/kg/day (table 4-5). This indicates a good animal tolerability in treatment with these compounds.

In conclusion, new imidazolidine compounds LPSF-FZ4 and LPSF-PT5 such as PZQ demonstrated to low plasma lipids levels. It is request furthermore studies for better comprehension of effects on lipid metabolism. LPSF-FZ4 showed to be a potential candidate for hypotriglyceridemic therapy who could be aim at combat cardiovascular diseases which are main causes of death in world [52].

4.6. Acknowledgements

This work was supported by CNPQ, FINEP and CAPES.

4.7. References

- [1] Chitsulo L, Engels D, Montresor A, Savioli L. The global status of schistosomiasis and its control. *Acta Trop* 2000; 77:41-51.
- [2] Blanchard TJ. Schistosomiasis. *Travel Med Infect Dis* 2004; 2: 5-11.

- [3] Ferrari MLA, Coelho MPZ. Efficacy of oxamniquine and praziquantel in the treatment of Schistosoma mansoni infection: a controlled trial. *Bulletin of the World Health Organization* 2003; 81:3.
- [4] Johnston DA. Opening the con of worms: molecular analysis of Schistosomiasis mansoni population. *Parasitol Today* 1993; 9: 286-291.
- [5] Coelho RB. Anatomia Patológica das Afecções Hepáticas, Imprensa Universitária, Universidade Federal de Pernambuco: Recife, 1971.
- [6] Mansour MM, Farid Z, Bassily S, Salah L, Watten R. Serum enzyme tests in hepatosplenomegaly schistosomiasis. *Trans R Soc Trop Med Hyg* 1982; 76(Suppl 1): 109-111.
- [7] Coutinho-Abath E, Barbosa JM, Amaral JA. Alterações bioquímicas na esquistossomose mansônica humana, como especial referência ao metabolismo lipídico. *J Bras Med* 1966; 11: 157-168.
- [8] Gillett MPT, Carvalho VC. Schistosoma mansoni: a comparative study of plasma and erythrocyte lipid alterations in the experimentally infected mouse and in selected human patients. *Exp Parasitol* 1978; 44: 173-180.
- [9] Owen J, Costa J, Carvalho V, Gillett M. Effect of experimental schistosomiasis mansoni on plasma and erythrocyte lipids and on plasma lecithin: cholesterol acyltransferase in the mouse. *Rev Bras Biol* 1978; 38(Suppl 4): 913-918.
- [10] Vieira E, De Oliveira D, Dimenstein R, Gillett M. Mouse plasma phospholipid metabolism: contributions of lecithin: cholesterol acyltransferase, phospholipase A and lysophospholipase activities. *Arq. Biol. Tecnol* 1992; 35(Suppl 3): 517-529.
- [11] Lima V, Sena V, Stewart B, Owen J, Dolphin P. An evaluation of *Callithrix jacchus* (saguí) as an experimental model for the dyslipoproteinemia of human schistosomiasis mansoni. *Biochim Biophys Acta* 1998; 1393(Suppl 2):235-243.
- [12] Caffrey CR. Chemotherapy of schistosomiasis: present and future. *Curr Opin Chem Biol* 2007; 11: 433-439.
- [13] Doenhoff MJ, Pica-Mattoccia L. Praziquantel for the treatment of schistosomiasis: its use for control in areas with endemic disease and prospects for drug resistance. *Expert Rev Anti Infect Ther* 2006; 4: 199-210.
- [14] Fallon PG, Doenhoff MJ. Drug-resistant schistosomiasis: resistance to praziquantel and oxamniquine induced in *Schistosoma mansoni* in mice is drug specific. *Am J Trop Med Hyg* 1994; 51: 83-88.

- [15] Stelma F, Talla I, Sow S, Kongs A, Niang M, Polman K, Deelder AM, Grysell B. Efficacy and side effects of praziquantel in an epidemic focus of *Schistosoma mansoni*. Am J Trop Med Hyg 1995;53(Suppl 2):167-70
- [16] Xiao S, Keiser J, Chollet J, Utzinger J, Dong Y, Endriss Y, Vennerstrom JL, Tanner M. In vitro and in vivo activities of synthetic trioxolanes against major human schistosome. Antimicrob Agents Chemother 2007; 51:1440-1445.
- [17] Moreira L, Pilo-Veloso D, De Mello R, Coelho P, Nelson D. A study of the activity of 2-(alkylamino)-1-phenyl-1-ethanethiosulfuric acids against infection by *Schistosoma mansoni* in a murine model. Trans R Soc Trop Med Hyg. 2007; 101: 385-390.
- [18] Barreiro EJ, Fraga CAM. Química medicinal: as bases moleculares da ação dos fármacos, Artmed Editora: Porto Alegre, 2001.
- [19] Thenmozhiyal JC, Wong PTH, Echui WK. Anticonvulsant activity of phenylmethylenehydantoins: a structure-activity relationship. J Med Chem 2004; 47:1527-1535.
- [20] Dylag T, Zygmunt M, Dorota M, Handzlik J, Bednarski M, Filipek B, Kieć-Kononowicz K. Synthesis and evaluation of in vivo activity of diphenylhydantoin basic derivatives. Eur J Med Chem 2004; 39:1013-1027.
- [21] Robinson RP, Laird E, Donahue K, Lopresti-Morrow L, Mitchell P, Reese M, Reeves LM, Rouch A, Stam EJ, Yocom SA. Design and synthesis of 2-Oxo-imidazolidine-4-carboxylic acid hydroxyamides as potent matrix metalloproteinase-13Inhibitors. Bioorg Med Chem Lett 2001; 11:1211-1213.
- [22] Pitta MGR, Silva ACA, Neves JKAL; Silva PG, Irmão JI, Malagueño E, Santana JV, Lima MCA, Galdino SL, Pitta IR, Albuquerque MCPA. New imidazolidinic bioisosters: potential candidates for antischistosomal drugs. Mem Inst Oswaldo Cruz 2006; 101(Suppl 5):313-316.
- [23] Albuquerque MCPA, Silva TG, Pitta MGR, Silva ACA, Malagueño E, Santana JV, Lima MCA, Galdino SL, Barbe J, Pitta IR. Synthesis and schistosomicidal activity of new substituted thioxo-imidazoline compounds. Pharmazie 2005; 60(Suppl 1):13-17.
- [24] Oliveira SM, Albuquerque MCPA, Pitta MGR, Malagueño E, Santana JV, Lima MCA, Pitta IR, Galdino SL. A resposta do *Schistosoma mansoni* mantido in vitro frente a derivados imidazolidinônicos. Acta Farmaceutica Bonaerense 2004; 23(Suppl 3):243-248.
- [25] Fletcher EJ, Kirsten E, Beech J. Increases specific triacylglycerol fatty esters in skeletal muscle from horses with hyperkalemic periodic paralysis. Biochim Biophys Acta - Lipids and Lipid Metabolism 1993; 1168 (Suppl 3):292-298.
- [26] Wierzbicki A. Imidazole derivatives as cholesterol-lowering agents. Int J Cardiol 2003;90:145-146.

- [27] Shamkhany K, Azarpira M, AKBAR M. An open label crossover trial of effects of metronidazol on hyperlipidaemia. *Int J Cardiol* 2003;90:143-146.
- [28] Goerdt C, Keith M, Rubins HB. Effects of phenytoin on plasma high-density lipoprotein cholesterol levels in men with low levels of high-density lipoprotein cholesterol. *J Clin Pharmacol* 1995; 35:767-775.
- [29] Trocho C, Escolà-Gil J, Ribas V, Benítez S, Martín-Campos J, Rotllan N, Osaba L, Ordóñez-Llanos J, González-Sastre F, Blanco-Vaca F. Phenytoin treatment reduces atherosclerosis in mice through mechanisms independent of plasma HDL-cholesterol concentration. *Atherosclerosis* 2004; 174:275-285.
- [30] Dubin M, Grinblat L, Fernandez Villamil S, Stoppani A. Nitrofuran inhibition of microsomal lipid peroxidation. *FEBS Lett* 1987; 220 (Suppl 1):197-200.
- [31] Houston M, Burger C, Hays J, Nadeau J, Swift L, Bradley C A, Olafsson L. The effects of clonidine hydrochloride versus atenolol monotherapy on serum lipids, lipid subfractions, and apolipoproteins in mild hypertension. *Am Heart J* 1990; 120 (Suppl 1):172-179.
- [32] Lima M, Costa L, Góes A, Galdino S, Pitta I, Luu-Doc C. Synthèse et activité antimicrobienne des chlorobenzyl benzylidène imidazolidinediones et thiazolidinediones substituées. *Pharmazie* 1992; 4(Suppl 3):182-184.
- [33] Brandão SSF, Rocha Filho JA, Chantegrel J, Albuquerque JFC, Ximenes EA, Galdino SL, Pitta IR, Perrissin M, Luu-Duc C. Synthèse et structure des arylazo-imidazolidines et arylidènethiazolidines substituées. *Ann Pharmaceutiques Françaises* 1997; 55 (Suppl 5):206-211.
- [34] Duvall RH, Dewitt WB. An improved perfusion technique for recovering adult schistosomes from laboratory animals. *Am J Trop Med Hyg* 1967; 16(Suppl 4):483-486.
- [35] Malagenõ E, Santana J. Etiologia. In: Malta J, editors. *Esquistossomose Mansônica*. Recife: Editora Universitária, 1994:26-46.
- [36] Bogliolo L. Anatomical picture of liver in hepatosplenic schistosomiasis mansoni. *Ann Trop Med Parasitol* 1957;51:1-14.
- [37] El-Lakkany NM, Seif el-Din SH, Badawy AA, Ebeid FA. Effect of artemether alone and in combination with grapefruit juice on hepatic drug-metabolising enzymes and biochemical aspects in experimental Schistosoma mansoni. *Int J Parasitol* 2004; 34:1405–1412.
- [38] Meyer F, Meyer H, Bueding E. Lipid metabolism in the parasitic and free-living flatworms, *Schistosoma mansoni* and *Dugesta dorotocephalia*. *Biochim Biophys Acta* 1970; 210: 257-266.

- [39] Bennett M, Cauldfild J. (1991). Specific binding of human low-density-lipoprotein to the surface of schistosomula of *Schistosoma mansoni* and ingestion by the parasite. Am J Pathol 1991;131:1173-1182.
- [40] Rumjanek FD, McLaren DJ, Smithers SR (1983). Serum-induced expression of a surface protein in schistosomula of *Schistosoma mansoni* - a possible receptor for lipid uptake. Mol Biochem Parasitol 1983; 9: 337-350
- [41] Alving CR, WASSEF NM. Naturally occurring antibodies to cholesterol: a new theory of LDL cholesterol metabolism. Immunol Today 1999;20:362-366.
- [42] Lima VLM, Coelho LCBB, Owen JS, Kennedy JF, Dolphin PJ. Lecithin: cholesterol acyltransferase as a plasma glycoprotein: an overview. Carbohydr Polym 2004; 55:179-191.
- [43] Sukkar MY, Omer AH, El Din Ahmed N. Impaired glucose tolerance in hepatic schistosomiasis. Trans R Soc Trop Med Hyg 1974; 68(Suppl 4):327-332.
- [44] Awadalla HN, el-Fiky R, Helmi AM. Letter: The pancreas and blood sugar changes in mice infected with *Schistosoma mansoni*. Trans R Soc Trop Med Hyg 1974;68(Suppl 5):410-411.
- [45] Doenhoff MJ, Stanley RG, Griffiths K, Jackson CL. An anti-atherogenic effect of *Schistosoma mansoni* infections in mice associated with a parasite-induced lowering of blood total cholesterol. Parasitol 2002; 125:415-421.
- [46] Malheiros S, Brito M, Brites D, Meirelles N. (2000). Membrane effects of trifluoperazine, dibucaine and praziquantel on human erythrocytes. Chem Biol Interact 2000;126:79-95.
- [47] Andrews P. Praziquantel: mechanisms of anti-schistosomal activity. Pharmacol Ther 1985;29:129-156.
- [48] Ahmed S, Smith J, Nicholls P, Whomsley R, Cariuk P.. Synthesis and biological evaluation of imidazole based compounds as cytochrome P-450 inhibitors. Drug Des Discov 1995; 13:27-41.
- [49] Burnett J, Wilcox L, Huff M. Acyl coenzyme A: cholesterol acyltransferase inhibition and hepatic apolipoprotein B secretion. Clin Chim Acta 1999; 286: 231-242.
- [50] Kempen H, Van Son K, Cohen L, Griffioen M, Verboom H, Havekes L. Effect of ketoconazole on cholesterol synthesis and on HMG-CoA reductase and LDL-receptor activities in Hep G2 cells. Biochem Pharmacol 1987;36:1245-1249.
- [51] Elokdah H, Sulkowski T, Cochran D, McKean ML, Quinet E. Effects of 2-(Substituted-sulfanyl)-3,5-dihydro-imidazole-4-one and 2-(Substituted-sulfanyl)-1H-imidazole-4,5-dione derivatives on Serum HDL-cholesterol. Bioorg Med Chem Lett 2000; 10:1791-1794.

[52] Reddy KS. Cardiovascular disease in non-western countries. *N Engl J Med* 2004; 350:2438-2440.

4.8. Tables

Table 1 - Comparison of biochemical parameters from uninfected and infected mice groups before treatment.

Parameters	Groups		Significance
	Uninfected	Infected	
Total Cholesterol (mmol/l)	2.14 ± 0.04	1.75 ± 0.03	p < 0.001
Triglycerides (mmol/l)	1.48 ± 0.05	1.12 ± 0.03	p < 0.001
Total Phospholipids (mmol/l)	12.60 ± 0.26	12.10 ± 0.25	p > 0.05
Aspartate Aminotransferase (U/l)	32.43 ± 0.79	45.41 ± 1.67	p < 0.001
Alanine Aminotransferase (U/l)	15.21 ± 0.35	24.75 ± 1.34	p < 0.001

Statistical significance determined by unpaired t test. Data are stated as means ± S.E.M

Table 2 - Plasma Total Cholesterol levels before and after 15 days of treatment with praziquantel (PZQ), 3-benzyl-5-(4-chloro-arylazo)-4-thioxo-imidazolidin-2-one (LPSF-PT5) and 3-(4-chloro-benzyl)-5-(4nitro-benzylidene)-imidazolidine-2,4-dione (LPSF-FZ4) in schistosomiasis mansoni infected and uninfected mice.

Groups (n = 8)	Concentration (mg/kg/day)	Plasma Total Cholesterol (mmol/l)							
		CMC		PZQ		LPSF-PT5		LPSF-FZ4	
		Before	After	Before	After	Before	After	Before	After
Infected	50	1.81 ± 0.10	1.61 ± 0.09	1.74 ± 0.09	1.58 ± 0.07	1.79 ± 0.06	1.55 ± 0.09	1.73 ± 0.12	1.49 ± 0.08
Infected	100	1.74 ± 0.05	1.55 ± 0.07	1.83 ± 0.05	1.38 ± 0.06***	1.72 ± 0.09	1.40 ± 0.05**	1.70 ± 0.07	1.58 ± 0.05
Uninfected	50	2.19 ± 0.12	2.19 ± 0.08	2.22 ± 0.18	2.25 ± 0.30	2.25 ± 0.12	2.22 ± 0.07	2.28 ± 0.11	2.26 ± 0.11
Uninfected	100	2.05 ± 0.13	2.10 ± 0.18	2.00 ± 0.07	1.75 ± 0.05	2.09 ± 0.06	1.92 ± 0.05	2.02 ± 0.09	2.08 ± 0.08

Data are stated as means ± S.E.M. **p<0.01; ***p<0.001 when compared simultaneously all compounds (Two-way ANOVA followed by Bonfferroni test) before and after treatment for each concentration.

Table 3 - Plasma triglycerides levels before and after 15 days of treatment with praziquantel (PZQ), 3-benzyl-5-(4-chloro-arylazo)-4-thioxo-imidazolidin-2-one (LPSF-PT5) and 3-(4-chloro-benzyl)-5-(4nitro-benzylidene)-imidazolidine-2,4-dione (LPSF-FZ4) in schistosomiasis mansoni infected and uninfected mice.

Groups (n = 8)	Concentration (mg/kg/day)	Plasma Triglycerides (mmol/l)							
		CMC		PZQ		LPSF-PT5		LPSF-FZ4	
		Before	After	Before	After	Before	After	Before	After
Infected	50	1.14 ± 0.08	0.95 ± 0.07	1.02 ± 0.11	0.86 ± 0.10	1.14 ± 0.12	0.86 ± 0.09	1.07 ± 0.10	0.59 ± 0.04 ***
Infected	100	1.13 ± 0.06	0.91 ± 0.07	1.20 ± 0.04	0.82 ± 0.10**	1.11 ± 0.02	0.76 ± 0.05**	1.14 ± 0.11	0.68 ± 0.05***
Uninfected	50	1.37 ± 0.12	1.45 ± 0.30	1.35 ± 0.22	1.01 ± 0.10	1.66 ± 0.21	1.42 ± 0.16	1.53 ± 0.20	1.12 ± 0.06*
Uninfected	100	1.50 ± 0.11	1.59 ± 0.15	1.57 ± 0.13	1.16 ± 0.16	1.51 ± 0.09	1.32 ± 0.15	1.52 ± 0.14	1.19 ± 0.06*

Data are stated as means ± S.E.M.. *p < 0.05; **p<0.01; ***p<0.001 when compared simultaneously all compounds (Two-way ANOVA followed by Bonfferroni test) before and after treatment for each concentration.

Table 4 - Plasma aspartate aminotransferase activity before and after 15 days of treatment with praziquantel (PZQ), 3-benzyl-5-(4-chloro-arylazo)-4-thioxo-imidazolidin-2-one (LPSF-PT5) and 3-(4-chloro-benzyl)-5-(4nitro-benzylidene)-imidazolidine-2,4-dione (LPSF-FZ4) in schistosomiasis mansoni infected and uninfected mice.

Groups (n = 8)	Concentration (mg/kg/day)	Aspartate Aminotransferase (U/l)							
		CMC		PZQ		LPSF-PT5		LPSF-FZ4	
		Before	After	Before	After	Before	After	Before	After
Infected	50	39.56 ± 1.72	35.66 ± 2.80	41.25 ± 2.30	35.85 ± 1.83	41.10 ± 1.67	36.23 ± 2.86	43.74 ± 1.82	38.33 ± 2.24
Infected	100	51.47 ± 2.25	49.15 ± 2.19	56.62 ± 2.66	79.79 ± 1.85**	54.58 ± 2.06	59.42 ± 2.26	42.08 ± 2.46	36.93 ± 1.63
Uninfected	50	31.26 ± 1.96	32.36 ± 2.11	31.56 ± 2.37	27.97 ± 1.46	31.71 ± 1.85	25.98 ± 2.66	30.08 ± 1.93	26.66 ± 2.69
Uninfected	100	34.76 ± 2.08	33.75 ± 2.47	37.49 ± 1.66	51.66 ± 2.55**	30.84 ± 2.04	34.46 ± 2.06	33.60 ± 2.03	28.10 ± 2.02

Data are stated as means ± S.E.M. **p<0.01 when compared simultaneously all compounds (Two-way ANOVA followed by Bonferroni test) before and after treatment for each concentration.

Table 5 - Plasma alanine aminotransferase activities before and after 15 days of treatment with praziquantel (PZQ), 3-benzyl-5-(4-chloro-arylazo)-4-thioxo-imidazolidin-2-one (LPSF-PT5) and 3-(4-chloro-benzyl)-5-(4nitro-benzylidene)-imidazolidine-2,4-dione (LPSF-FZ4) in schistosomiasis mansoni infected and uninfected mice.

Groups (n = 8)	Concentration (mg/kg/day)	Alanine Aminotransferase(U/I)							
		CMC		PZQ		LPSF-PT5		LPSF-FZ4	
		Before	After	Before	After	Before	After	Before	After
Infected	50	23.06 ± 2.58	23.59 ± 2.62	22.17 ± 1.97	18.91 ± 1.56	25.57 ± 2.66	19.10 ± 2.56	26.82 ± 1.99	21.24 ± 2.84
Infected	100	20.43 ± 2.87	20.71 ± 2.07	18.80 ± 1.72	21.27 ± 2.61	21.55 ± 2.26	23.52 ± 1.76	25.08 ± 1.79	20.99 ± 2.87
Uninfected	50	14.58 ± 1.11	13.00 ± 0.75	14.95 ± 0.72	12.32 ± 0.89	15.28 ± 0.97	11.31 ± 0.84	14.04 ± 0.96	10.45 ± 0.68
Uninfected	100	16.21 ± 0.46	15.50 ± 0.41	15.51 ± 1.20	16.93 ± 1.38	15.79 ± 1.28	15.02 ± 1.17	14.68 ± 0.70	12.36 ± 1.62

Data are stated as means ± S.E.M.

4.9. Figure legends

Figure 1 – Chemical structure of compounds, 3-(4-chloro-benzyl)-5-(4nitro-benzylidene)-imidazolidine-2,4-dione (LPSF-FZ4) (A) and 3-benzyl-5-(4-chloro-arylazo)-4-thioxo-imidazolidin-2-one (LPSF-PT5) (B).

5. CONCLUSÕES

- I. A infecção por *S. mansoni* (Cepa BH) promoveu diminuição nos níveis de colesterol total e triglicerídeos em camundongos. Entretanto não foi observada redução nos níveis de fosfolipídios totais plasmáticos. A função hepática dos camundongos infectados por *S. mansoni* (Cepa BH) mostrou-se alterada, em detrimento das elevações de atividades enzimáticas de aspartato aminotransferase e alanina aminotransferase.
- II. O tratamento com o derivado imidazolidínico LPSF-PT5 e Praziquantel, a uma concentração de 100mg/kg/dia promoveu redução significativa de colesterol total e triglicerídeos plasmáticos em camundongos com a esquistossomose além de possuir boa tolerabilidade.
- III. O derivado imidazolidínico LPSF-FZ4, a uma concentração de 50 mg/kg/dia e 100 mg/kg/dia, mostrou apresentar potencial atividade hipolipidêmica, reduzindo os triglicerídeos plasmáticos em camundongos infectados e não infectados, além de ser bem tolerado.

6. ANEXOS

6.1. Normas para redação de artigos para a revista “Parasitology International”

PARASITOLOGY INTERNATIONAL

The Official, International Journal of the [Japanese Society of Parasitology](#)

Guide for Authors

[Guide for Authors click for pdf file](#)

Submission of a paper to *Parasitology International*, including a revised version, implies the transfer of copyright from the author(s) to the publisher and therefore it is imperative that the corresponding author has obtained the approval of all other authors to the text and that it does not contain information previously published (except as a meeting abstract or by submission of a sequence data to an electronic database) and is not under consideration for publication elsewhere. Publication in *Parasitology International* is taken to imply the authors' willingness to comply with reasonable requests to supply reagents such as recombinant clones and monoclonal antibodies, and sequence data in electronic form to persons lacking access to computer databases.

Submission

Submission to this journal proceeds totally online. Use the following guidelines to prepare your article. Via the homepage of this journal ([↗](#) <http://www.elsevier.com/locate/parint>) you will be guided stepwise through the creation and uploading of the various files. The system automatically converts source files to a single Adobe Acrobat PDF version of the article, which is used in the peer-review process. Please note that even though manuscript source files are converted to PDF at submission for the review process, these source files are needed for further processing after acceptance. All correspondence, including notification of the Editor's decision and requests for revision, takes place by e-mail and via the Author's homepage, removing the need for a hard-copy paper trail.

The above represents a very brief outline of this form of submission. It can be advantageous to print this "Guide for Authors" section from the site for reference in the subsequent stages of article preparation.

Papers accepted for publication should be concise as possible. Please submit, with the manuscript, the names and addresses of three potential referees.

Upon acceptance of an article, authors will be asked to sign a "Journal Publishing Agreement" (for more information on this and copyright see [↗](#) <http://www.elsevier.com/copyright>). Acceptance of the agreement will ensure the widest possible dissemination of information. An e-mail (or letter) will be sent to the corresponding author confirming receipt of the manuscript together with a "Journal

"Publishing Agreement" form or a link to the online version of this agreement.

If excerpts from other copyrighted works are included, the author(s) must obtain written permission from the copyright owners and credit the source(s) in the article. Elsevier has pre-printed forms for use by authors in these cases: contact Elsevier's Rights Department, Oxford, UK: phone (+44) 1865 843830, fax (+44) 1865 853333, e-mail permissions@elsevier.com. Requests may also be completed online via the Elsevier homepage (<http://www.elsevier.com/locate/permissions>).

Submission of sequence data to databases

The easiest and preferred method of submitting sequence data is to use the Authorin programme, which is available free of charge for PC or Macintosh systems from GenBank, National Centre for Biotechnology Information, Bldg. 38A, Room 8N-803, 8600 Rockville Pike, Bethesda, MD 20894, USA (voice: +1 301 4962475; fax +1 3012 4809241; e-mail: authoin@ncbi.nlm.nih.gov). Files generated by Authorin may be sent by e-mail to gb-bub@ncbi.nlm.nih.gov or copied to floppy disk and mailed to GenBank. For each sequence, a unique accession number will be issued by the database (within 24 hours if received via e-mail). The accession number should be included as a footnote on the first page of the article: 'Note: Nucleotide sequences data reported in this paper are available in the EMBL, GenBank and DDJB data bases under the accession number(s) -.' If requested, GenBank will withhold release of data until the appearance of your paper. Updates, corrections or notification for publication should be sent to update@ncbi.nlm.nih.gov.

Instructions for authors regarding GenBank/DNA sequence linking

DNA sequences and GenBank Accession numbers. Many Elsevier journals cite "gene accession numbers" in their running text and footnotes. Gene accession numbers refer to genes or DNA sequences about which further information can be found in the databases at the National Center for Biotechnical Information (NCBI) at the National Library of Medicine. Elsevier authors wishing to enable other scientists to use the accession numbers cited in their papers via links to these sources, should type this information in the following manner:

For each and every accession number cited in an article, authors should type the accession number in bold, underlined text. Letters in the accession number should always be capitalised. (See example 1 below). This combination of letters and format will enable Elsevier's typesetters to recognize the relevant texts as accession numbers and add the required link to GenBank's sequences.

Example 1: "GenBank accession nos. AI631510 , AI631511 , AI632198, and BF223228), a B-cell tumor from a chronic lymphatic leukemia (GenBank accession no. BE675048), and a T-cell lymphoma (GenBank accession no. AA361117)".

Authors are encouraged to check accession numbers used very carefully. An error in a letter or number can result in a dead link.

In the final version of the printed article , accession number text will not appear bold or underlined (see Example 2 below).

Example 2: "GenBank accession nos. AI631510, AI631511, AI632198, and BF223228), a B-cell tumor from a chronic lymphatic leukemia (GenBank accession no. BE675048), and a T-cell lymphoma (GenBank accession no. AA361117)".

In the final version of the electronic copy , the accession number text will be linked to the appropriate source in the NCBI databases enabling readers to go directly to that source from the article (see Example 3 below).

Example 3: "GenBank accession nos. AI631510, AI631511, AI632198, and BF223228), a B-cell tumor from a chronic lymphatic leukemia (GenBank accession no. BE675048), and a T-cell lymphoma (GenBank accession no. AA361117)".

Special Subject Repositories

Certain repositories such as PubMed Central ("PMC") are authorized under special arrangement with Elsevier to process and post certain articles such as those funded by the National Institutes of Health under its Public Access policy (see  <http://www.elsevier.com> for more details on our policy).

Articles accepted for publication in an Elsevier journal from authors who have indicated that the underlying research reported in their articles was supported by an NIH grant will be sent by Elsevier to PMC for public access posting 12 months after final publication. The version of the article provided by Elsevier will include peer-review comments incorporated by the author into the article. Because the NIH "Public Access" policy is voluntary, authors may elect not to deposit such articles in PMC. If you wish to "opt out" and not deposit to PMC, you may indicate this by sending an e-mail to mail to: NIHauthorrequest@elsevier.com.

Manuscripts

Manuscripts should be in English. They should be divided into: (1) title page-include a succinct title (which should not normally exceed 100 characters and should not contain any subtitles or abbreviations), the names of the authors including a given name for each, the institutions with city, state and country where the work was performed, the name and complete address (including telephone, fax and e-mail) of the corresponding author, a list of abbreviations and a list of addresses of authors who have moved from the institution where the work was performed, (2) abstract-maximum 250 words (3) keywords (3-6 indexing terms), (4) introduction, (5) materials and methods, (6) results, (7) discussion, (8) acknowledgements (grant support and technical support to be listed here), (9) references, (10) tables and (11) figure legends. A recent issue of the journal should be consulted for details. In the interests of clarity and brevity, it may sometimes be advantageous to combine the results and discussion into a single section. Everyone makes minor modifications to standard methods. Do not describe standard materials and methods or modifications unless they have significant and demonstrable ability. Do not duplicate descriptions of methodology in the figure legends. Generic and species names should be typed out in full the first time mentioned - in the title, the summary and the text- and thereafter the generic name should be abbreviated. Words or letters to be printed in italics should either be in italics or underlined. The metric system should be used

throughout.

Research Notes

These are intended for the publication of brief definitive reports, primarily of complete DNA sequence data, methods, case reports that do not merit a full length publication. Maximum length is four printed pages, including one or two figures. Only the salient points of a long DNA sequence should be published, as the whole sequence will be available from a computer database. The title, authorship and affiliations will be in the standard format of the journal. The text should not be sectioned, except for references. Essential experimental details may be incorporated into a figure legend. To facilitate rapid publication, authors will be expected to supply high quality copy and expedite any necessary revisions, although decisions will normally be yes or no, based on the quality and appropriateness of the initial submission.

Minireviews.

Minireviews are initiated on topics of current, significant advancement in the field. The reviews should be short (maximum 4000 words), current, specific and potentially provocative. They should provide a balanced synthesis from the available data rather than a simple regurgitation of results, but not to be overly speculative. If possible, they should provide new concepts and ideas extending across different parasite systems. The text can be divided into simple sections with a succinct abstract. Minireviews will be subjected to the established review process, and published by an accelerated schedule if accepted. Minireviews should be submitted via EES. Please use the "Mini-Review" title of the submission menu to ensure that your article is published in the correct section of the journal.

References. In the text, references should be numbered singly in square brackets in order of their citation, e.g., [2,3,5-7]. In the list, references should be numbered in the order of citation in the text, not in alphabetical order. Unpublished data, personal communications and papers in preparation or 'submitted' should not be listed in the references (but may be incorporated at the appropriate place in the text); work 'in press' may be listed only if it has been accepted for publication. Personal communications must be accompanied by a letter from the named person(s) giving permission to quote such information. Abstracts (whether published or not), theses and similar material are not to be quoted in the list. If necessary, they can be referred to in the text in parentheses. Periodicals [1], books [2] and edited books [3] should accord with the following examples;

[1] Perrine KG, Denker JA, Nilsen TW. A multi-copy gene encodes a potentially protective antigen in *Brugia malayi*. *Mol Biochem Parasitol* 1988;30: 97-104.

[2] Davis LG, Dibner MD, Battey JF. *Basic Methods in Molecular Biology*, Elsevier: Amsterdam, 1986.

[3] Chang K-P, Fong D, Bray RS. *Biology of Leishmania and leishmaniasis*. In: Chang K-P, Bray RS, editors. *Leishmaniasis*. Amsterdam: Elsevier, 1985: 1-30.

[4] Lai AA, De la Cruz VF, Campbell GH, Procell PM, Collins WE, McCuan TF. Structure of the circumsporozoite gene of Plasmodium malrize. Mol Biochem Parasitol (in press).

Abbreviations of journal titles should conform to those adopted by the List of the Serial Title Word Abbreviations, ISDS International Centre 20, rue Bachaumont, 75002 Paris, France (ISBN 2-904938-02-8).

Tables.

Each table should be typed double-spaced on a separate sheet and have a short descriptive title. A legend may be placed under the table. Footnotes should be identified in the table by a, b, c, etc.

Figures.

Figures must be in a form and condition suitable for high quality reproduction. Lettering should be clear and of adequate size to be legible after reduction. Consider the printed page and column proportions when preparing figures. Multiple panels of a single figure must be mounted together. Each DNA sequence figure must fit on a single page. Place numbering at one end of each line, not on separate lines, and avoid excessive line spacing. Consider placing nucleotide and protein data in separate panels, using single-letter amino acid abbreviations for the protein sequence and grouping nucleotides either continuously or in blocks of ten separated by one space. Preferably use a sans-serif font. Upper case is standard, except that introns or other features can be usefully distinguished by lower case. Provide sharp laser-printer or imagesetter copy. Nucleotide sequences of long coding regions, where the amino acid sequence is the primary feature, and long DNA sequences may, at the editor's discretion, be omitted from the printed paper. They can be obtained from electronic databases or from the authors. If, together with your accepted article, you submit usable colour figures then Elsevier will ensure, at no additional charge, that these figures will appear in colour on the web (e.g., ScienceDirect and other sites) regardless of whether or not these illustrations are reproduced in colour in the printed version. For colour reproduction in print, you will receive information regarding the costs from Elsevier after receipt of your accepted article. Please note: Because of technical complications which can arise by converting colour figures to 'grey scale' (for the printed version should you not opt for colour in print) please submit in addition usable black and white prints corresponding to all the colour illustrations.

As only one figure caption may be used for both colour and black and white versions of figures, please ensure that the figure captions are meaningful for both versions, if applicable. Figure legends should be typed double spaced at the end of the text, not on the figures. Figures should be checked extremely carefully, particularly after revisions. No changes to figures will be possible after acceptance of the manuscript.

A detailed guide on electronic artwork is available on our website:  <http://www.elsevier.com/artworkinstructions>

Detailed Instructions.

Abbreviations, symbols, chemical and biochemical nomenclature, etc, should follow

the recommendations given in the Journal of Biological Chemistry (Vol. 268, pp. 14543-14551). Avoid abbreviations which are not in common use across the field of parasitology. Those used should be defined in the text on first usage and listed as a footnote on the title page. Do not introduce abbreviations unless they are used at least 4 times.

Proofreading. One set of page proofs in PDF format will be sent by email to the corresponding author to be checked for typesetting/editing. No changes in or additions to the accepted (and subsequently edited) manuscript will be allowed at this stage. Proofreading is solely your responsibility. Addenda in proofs will be printed only in exceptional cases, and only after approval by the editors. Elsevier will do everything possible to get your article corrected and published as quickly and accurately as possible. Therefore, it is important to ensure that all of your corrections are sent back in one communication. Subsequent corrections will not be possible, so please ensure your first sending is complete.

Off prints

The corresponding author, at no cost, will be provided with a PDF file of the article via e-mail or, alternatively, 25 free paper off prints. The PDF file is a watermarked version of the published article and includes a cover sheet with the journal cover image and a disclaimer outlining the terms and conditions of use.

Editors.

Editor-in-Chief: Naoki Arizono, Department of Medical Zoology, Kyoto Prefectural University of Medicine, Kawaramachi-Hirokoji, Kyoto 602-8566, Japan, Tel: +81-75-251-5325; Fax:+81-75-251-5328; E-mail: pieditor@koto.kpu-m.ac.jp

Regional Editor (Europe and Africa): Karl Hoffman, University of Cambridge, Tennis Court Road, Cambridge CB2 1QP, UK, E-mail: kfh24@cam.ac.uk

Regional Editor (Americas): Fidel Zavala, Department of Molecular Microbiology and Immunology, Bloomberg School of Public Health, John Hopkins University, 615 N.Wolfe Street, Baltimore, MD, USA, E-mail: fzavala@jhsph.edu

Managing Editor : Tatsuya Tegoshi, Department of Medical Zoology, Kyoto Prefectural University of Medicine, Kawaramachi-Hirokoji, Kyoto 602-8566, Japan, Tel: +81-75-251-5325; Fax:+81-75-251-5328; Email: pieditor@koto.kpu-m.ac.jp

Language Polishing

For authors who require information about language editing and copyediting services pre- and post-submission please visit  <http://www.elsevier.com/wps/find/authorshome.authors/languagepolishing> or contact authorsupport@elsevier.com for more information. Please note Elsevier neither endorses nor takes responsibility for any products, goods or services offered by outside vendors through our services or in any advertising. For more information please refer to our Terms and Conditions  http://www.elsevier.com/wps/find/termsconditions.cws_home/termsconditions

Authors' rights

As an author you (or your employer or institution) retain certain rights; for details you are referred to: ↗<http://www.elsevier.com/wps/find/authorshome.authors/authorsrights>.

Author enquiries.

Authors can track accepted articles at ↗<http://www.elsevier.com/trackarticle> and set up e-mail alerts to inform them of when an article's status has changed, as well as copyright information, frequently asked questions and more.

Full details of electronic submission can be obtained from ↗<http://www.elsevier.com/locate/parint>Do not introduce abbreviations unless they are used at least 4 times.

Proofreading. One set of page proofs in PDF format will be sent by email to the corresponding author to be checked for typesetting/editing. No changes in or additions to the accepted (and subsequently edited) manuscript will be allowed at this stage. Proofreading is solely your responsibility. Addenda in proofs will be printed only in exceptional cases, and only after approval by the editors. Elsevier will do everything possible to get your article corrected and published as quickly and accurately as possible. **Therefore, it is important to ensure that all of your corrections are sent back in one communication.** Subsequent corrections will not be possible, so please ensure your first sending is complete.

Reprints. The author will receive an order form with the proofs on which reprints above the 25 free per contribution may be ordered.

Editors.

Editor-in-Chief: Naoki Arizono, Department of Medical Zoology, Kyoto Prefectural University of Medicine, Kawaramachi-Hirokoji, Kyoto 602-8566, Japan, Tel: +81-75-251-5325; Fax:+81-75-251-5328; Email: pieditor@koto.kpu-m.ac.jp

Regional Editor (Europe): Prof. R. Carter, University of Edinburgh, School of Biological Sciences, Ashworth Laboratories, West Mains Road, Edinburgh EH9 3JT, U.K.; Tel: (131) 650 5558, Fax: (131) 668 3861, E-mail: r.carter@ed.ac.uk

Managing Editor : Tatsuya Tegoshi, Department of Medical Zoology, Kyoto Prefectural University of Medicine, Kawaramachi-Hirokoji, Kyoto 602-8566, Japan, Tel: +81-75-251-5325; Fax:+81-75-251-5328; Email: pieditor@koto.kpu-m.ac.jp

Language Polishing

For authors who require information about language editing and copyediting services pre- and post-submission please visit ↗<http://www.elsevier.com/wps/find/authorshome.authors/languagepolishing> or contact authorsupport@elsevier.com for more information. Please note Elsevier neither endorses nor takes responsibility for any products, goods or services offered by outside vendors through our services or in any advertising. For more information please refer

to our Terms and Conditions ↗
http://www.elsevier.com/wps/find/termsconditions.cws_home/termsconditions

Authors' rights

As an author you (or your employer or institution) may do the following: - make copies (print or electronic) of the article for your own personal use, including for your own classroom teaching use

- make copies and distribute such copies (including through e-mail) of the article to research colleagues, for the personal use by such colleagues (but not commercially or systematically, e.g., via an e-mail list or list server)
- post a pre-print version of the article on Internet websites including electronic pre-print servers, and to retain indefinitely such version on such servers or sites
- post a revised personal version of the final text of the article (to reflect changes made in the peer review and editing process) on your personal or institutional website or server, with a link to the journal homepage (on elsevier.com)
- present the article at a meeting or conference and to distribute copies of the article to the delegates attending such a meeting
- for your employer, if the article is a 'work for hire', made within the scope of your employment, your employer may use all or part of the information in the article for other intra-company use (e.g., training)
- retain patent and trademark rights and rights to any processes or procedure described in the article
- include the article in full or in part in a thesis or dissertation (provided that this is not to be published commercially)
- use the article or any part thereof in a printed compilation of your works, such as collected writings or lecture notes (subsequent to publication of your article in the journal)
- prepare other derivative works, to extend the article into book-length form, or to otherwise re-use portions or excerpts in other works, with full acknowledgement of its original publication in the journal

Author enquiries. Authors can keep a track on the progress of their accepted article, and set up e-mail alerts informing them of changes to their manuscript's status, by using the "Track a Paper" feature of Elsevier's [Author Gateway](#).

Full details of electronic submission and formats can be obtained from ↗
<http://authors.elsevier.com>

6.2. Resumos apresentados

II Reunião Regional da FeSBE, 2007

EFEITO DO TRATAMENTO COM LPSF-PT5 SOBRE OS NÍVEIS DE COLESTEROL E TRIGLICERÍDEOS EM PLASMA DE CAMUNDONGOS INFECTADOS POR *SCHISTOSOMA MANSONI*.

Apolinário Segundo, J.O**¹; Araújo, T.G¹; Lima, M. C. A²; César, F. A³; Neves, J. K. A. L³; Albuquerque, M. C. P. A⁴; Galdino, S.L²; Lima, V.L.M¹;

XX Congresso Brasileiro de Parasitologia, 2007

EFEITO DO TRATAMENTO DE DERIVADO IMIDAZOLIDÍNICO E PRAZIQUANTEL SOBRE OS NÍVEIS DE COLESTEROL TOTAL E TRIGLICERÍDEOS PLASMÁTICOS DE CAMUNDONGOS INFECTADOS POR *SCHISTOSOMA MANSONI*

José O.A.Segundo, Cleideana B. Silva, Juliana K.A.L.Neves, Fernanda A. César, Maria C.A. Lima, Mônica C.P. Albuquerque, Ivan R. Pitta, Suely L. Galdino, Vera L.M. Lima.